



---

# The Deepening, Lengthening and Widening of Berths 203 to 205 at Pier 2 Container Terminal, Port of Durban, KwaZulu-Natal Province

---

## Environmental Monitoring Report

Winter 2021

GE38097

*September 2021*



People • Expertise • Excellence

---

## Environmental Monitoring Report, September 2021

---

Chapter	Page
Report Details	iv
Revision Status	v
List of Figures	v
List of Tables	vii
Abbreviations	x
Executive Summary	xi
1 Background	1
2 Project Location and Baseline Environment	2
2.1 Port of Durban	2
2.2 Geology	3
2.3 Site Location	3
2.4 Aspects of environmental monitoring	4
3 Water Quality	5
3.1 Introduction	5
3.2 Methodology	7
3.3 Results	8
3.4 Conclusion	10

---

<b>4</b>	<b>Sediments</b>	<b>11</b>
4.1	Introduction	11
4.2	Methodology	14
4.3	Results	17
4.4	Conclusion	19
<b>5</b>	<b>Benthic Macrofauna</b>	<b>20</b>
5.1	Introduction	20
5.2	Methodology	21
5.3	Results	23
5.4	Conclusion	29
<b>6</b>	<b>Benthic Microalgae (Microphytobenthos)</b>	<b>30</b>
6.1	Introduction	30
6.2	Methodology	31
6.3	Results	32
6.4	Conclusion	34
<b>7</b>	<b>Biomonitoring using Mussels (<i>Perna perna</i>)</b>	<b>35</b>
7.1	Introduction	35
7.2	Methodology	37
7.3	Results	39
7.4	Conclusion	41
<b>8</b>	<b>Fish</b>	<b>42</b>

---

	8.1	Introduction	42
	8.2	Methodology	43
	8.3	Results	45
	8.4	Conclusion	48
9		<b>Birds</b>	<b>50</b>
	9.1	Introduction	50
	9.2	Methodology	50
	9.3	Results	51
	9.4	Conclusion	58
10		<b>General Conclusion</b>	<b>60</b>
	10.1	Water quality	60
	10.2	Sediment characteristics	60
	10.3	Benthic macrofauna	60
	10.4	Benthic microalgae	61
	10.5	Biomonitoring using mussels ( <i>Perna perna</i> )	61
	10.6	Fish	61
	10.7	Birds	61
11		<b>References</b>	<b>63</b>
		<b>Document Control and Disclaimer</b>	<b>72</b>

## Report Details

1	<b>Project name</b>	The Deepening, Lengthening and Widening of Berths 203 to 205 at Pier 2 Container Terminal, Port of Durban, KwaZulu-Natal Province
2	<b>Report title</b>	Environmental Monitoring Report, Winter 2021
3	<b>Authority reference</b>	No: 14/12/16/3/3/2/275
4	<b>Report status</b>	Draft 1
5	<b>Client</b>	Transnet National Ports Authority
6	<b>Scope</b>	Conduct quarterly monitoring during the two-year construction period of the Sandbank Extension. Thereafter, the monitoring should take once every six months during the construction of the Berth 203 to 205 Expansion
7	<b>Standard:</b>	Environmental Authorisation (NEAS: DEA/EIA/0000988/2012. DEA Ref: 14/12/16/3/3/32/275) and Central Sandbank Mitigation Plan (CSMP), October 2017
8	<b>Objective</b>	Conduct environmental monitoring, to track the health of the Durban bay over the construction period of the Berth 203 to 205 expansion project
9	<b>Methodology</b>	As per the CSMP Rev 06 Report, October 2017
10	<b>Date of fieldwork</b>	1, 10, 17, 23, 24 July 2021
11	<b>Dates of bird fieldwork</b>	Sampling done on the first spring tide of every month. This report includes bird results from December 2019 – May 2021, with the exception of April – July 2020.
12	<b>Report publish date</b>	10 September 2021
13	<b>Key notes</b>	Construction had not yet commenced at the time of this sample run
14	<b>Activity schedule</b>	<p><b><u>Quarterly Monitoring Events:</u></b></p> <ul style="list-style-type: none"> <li>• October 2018 (Spring I) – Sampling undertaken 9 - 18 October 2018</li> <li>• January 2019 (Summer I) – Did not occur, project put on hold.</li> <li>• April 2019 (Autumn I) – Did not occur, project put on hold.</li> <li>• August 2019 (Winter I) – Sampling undertaken 15 - 30 August 2019</li> <li>• October 2019 (Spring II) – Sampling undertaken 13 - 29 October 2019</li> <li>• January 2020 (Summer II) – Sampling undertaken 03 - 24 January 2020</li> <li>• June 2020 (Autumn II) – Sampling undertaken 05 – 23 June 2020. Note that autumn sampling performed in June due to National Lockdown.</li> <li>• July 2020 (Winter II) – Sampling undertaken 05 – 22 July, bird sampling undertaken throughout August.</li> <li>• October 2020 (Spring III) – Sampling undertaken 01 – 28 October</li> <li>• January 2021 (Summer III) – Sampling undertaken 13 – 29 January</li> <li>• April 2021 (Autumn III) – Sampling undertaken 12 – 28 April</li> <li>• July 2021 (Winter III) – Sampling undertaken 1 – 24 July</li> </ul>
15	<b>Authors</b>	<ul style="list-style-type: none"> <li>• Kolawole Akinoshun (PhD Candidate Marine Biology (Genetics), UKZN).</li> </ul>

		<ul style="list-style-type: none"> <li>• Charl Kruger (BSc Hons Environmental Management), GIBB Environmental (Environmental Scientist)</li> <li>• Robyn Phillips (PrSciNat, MSc Zoology) GIBB Environmental (Avifaunal Specialist)</li> <li>• Dr Lisa Skein (PhD Zoology), GIBB Environmental (Marine Scientist)</li> </ul> <b>Reviewers:</b> <ul style="list-style-type: none"> <li>• Dr Lisa Skein (PhD Zoology), GIBB Environmental (Marine Scientist)</li> <li>• Walter Fyvie (PrSciNat, MSc Environmental Biotechnology), GIBB Environmental (Senior Associate)</li> </ul>
16	<b>Contact person</b>	Charl Kruger Tel: +27 31 267 8572 Fax: +27 31 266 3310 Email: ckruger@gibb.co.za

## Revision Status

Rev No.	Issue Date	Revision Description
0	10/09/2021	Draft 1 for client review

## List of Figures

Figure 1: Ariel view of intertidal sandbank habitats (Northern Bank, Centre Bank, Little Lagoon, Island View, Bayhead Mangroves) in the Port of Durban (Google Earth, 2015).....3

Figure 2: Water quality monitoring sites (white pins) in the Port of Durban. ....7

Figure 3: Intertidal sediment sample sites in the Port of Durban. This includes 21 intertidal stations on the existing central sandbank (yellow pins – Int 10-31), 10 intertidal northern bank stations (yellow pins, Int 40-49), and additional 12 intertidal impact-monitoring stations on the central (black pins, Int 2 - 9). Sites immediately adjacent to the newly established sand bank (red pins, Int 53 - 63) will be sampled once development has commenced. Impact monitoring stations: Int2 – Int31, control monitoring stations: Int40 – Int49. Samples are collected at these sites for sediment characteristics, microalgae and benthic macrofauna. The area highlighted in red is where dredging is to take place.....15

Figure 4: Subtidal sites within the Port of Durban where sediment samples are taken. These sites represent three habitats: the central bank (Sub13 – Sub26), the little lagoon (Sub27 – Sub33), and the northern bank (Sub40 – Sub48). Impact monitoring stations: Sub13-Sub33, control stations: Sub40-Sub48. Samples are collected at these sites for sediment

---

characteristics, microalgae and benthic macrofauna. The area highlighted in red is where dredging is to take place.....	16
Figure 5: Mean ( $\pm$ standard error) (a) species richness, (b) abundance and (c) diversity of intertidal macrofaunal communities of the central and northern sandbanks. ....	24
Figure 6: Percentage abundance of various macrofaunal classes in communities on the central bank and northern bank.....	25
Figure 7: Mean ( $\pm$ standard error) (a) abundance, (b) species richness and (c) diversity of subtidal macrofaunal communities of the central bank, northern bank and little lagoon. ....	27
Figure 8: Percentage abundance of subtidal macrofauna classes in communities of the central bank, little lagoon and northern bank.....	28
Figure 9: Holes made by common sandprawn <i>Callichirus kraussi</i> through a plastic bag. (January 2021) .....	29
Figure 10: Chlorophyll-a concentrations ( $\mu\text{g}/\text{m}^2$ ) in 39 intertidal monitoring stations in the Port of Durban, winter 2021. Impact sites = Int2 – Int31 (central bank), control sites = Int40 – Int49 (northern bank). Red dotted lines: 20%tile (0.6 $\mu\text{g}/\text{m}^2$ ) and 80%tile (21.6 $\mu\text{g}/\text{m}^2$ ) intertidal winter threshold limits stipulated in the CSMP.....	33
Figure 11: Chlorophyll-a concentrations ( $\mu\text{g}/\text{m}^2$ ) in 25 subtidal monitoring stations in the Port of Durban, winter 2021. Impact sites = Sub13 – Sub34, control sites = Sub40 – Sub48. Red dotted lines: 20%tile (1.1 $\mu\text{g}/\text{m}^2$ ) and 80%tile (8.3 $\mu\text{g}/\text{m}^2$ ) subtidal winter threshold limits stipulated in the CSMP. ....	33
Figure 12: Locations of channel buoys for brown mussel ( <i>Perna perna</i> ) biomonitoring in the Port of Durban. Impact sites: 11, 12, 14, 16, 18 – 21; Control sites: 8 – 10, 13, 15, 17, 22 and 23.....	38
Figure 13: Median mercury concentrations (mg/kg) quantified from October 2020 (spring) to July 2021 (winter). Red line = baseline threshold value (0.29306 mg/kg). ....	41
Figure 14: Location of fish sampling stations in the Port of Durban. Yellow pins = control sites, green pins = impact sites, black pins = additional sampling sites that will be included after the construction of the berths to monitor sandbank recovery.....	44
Figure 15: Relative abundance (%) of fish species caught overall (all sites) and at impact and control sites. Note that the y-axis starts at 88%. ....	46
Figure 16: Relative mass (%) of fish species caught overall (all sites) and at impact and control sites.....	47

---

Figure 17: Aerial view of Durban Bay (Port of Durban) with 20 bird survey areas. ....	51
Figure 18: Accumulating plastic pollution on the Central Bank where monthly bird counts are conducted.....	53
Figure 19: Taxonomic composition (number and percentage of species) of bird communities recorded during monthly counts from June - August 2021.....	54
Figure 20: Composition of feeding guilds (number and percentage of species) recorded during monthly counts from June - August 2021.....	54
Figure 21: Total bird abundance per habitat recorded during monthly counts from June – August 2021. ....	56
Figure 22: Monthly bird abundance per habitat recorded from June – August 2021. ....	57

## List of Tables

Table 1: Water quality results of surface (< 1 m depth) and bottom layers recorded during the winter 2021 sampling event. All results given as mean $\pm$ standard deviation. ....	9
Table 2: Sediment grain size characterization. ....	11
Table 3: Total Organic Carbon (TOC) classification levels in sediments (adapted from Forbes & Demetriades, 2003). ....	12
Table 4: South African and NOAA sediment quality guidelines for trace element concentrations. Concentrations are parts per million (ppm) dry weight (mg/kg), ERL = Effects Range Low, ERM = Effects Range Median (DEA, n.d). ....	13
Table 5: Baseline threshold (warning) levels for sediment parameters (%sand, mud %, Total Organic Carbon (TOC) %, and trace metal concentrations) in intertidal and subtidal sediment samples from control and impact monitoring stations across the Port of Durban (adapted from Clark et al., 2017). Values for grain sizes (sand, mud) must fall between 10 – 90 <sup>th</sup> percentiles whereas values for trace metals must fall below the 90 <sup>th</sup> percentile. ....	14
Table 6: Key results of intertidal sediment characteristics (grain size, TOC, trace metal concentrations) in relation to the 90th percentile threshold limits of the CSMP. Values for grain sizes (sand, mud) must fall between 10 – 90th percentiles whereas values for trace metals must fall below the 90 <sup>th</sup> percentile. Values printed in red are outside of allowable ranges. Sites are grouped as overall (all sites), impact sites (Int3 – Int 31) and control sites (Int40 – Int49). Note that mercury concentrations are reported as < 0.03 ppm as a result of concentrations being lower than laboratory detection limits. ....	18



---

Table 7: Key results of subtidal sediment characteristics (grain size, TOC, trace metal concentrations) in relation to the 90th percentile threshold limits of the CSMP. Values for grain sizes (sand, mud) must fall between 10 – 90<sup>th</sup> percentiles whereas values for trace metals must fall below the 90<sup>th</sup> percentile. Values printed in red are outside of allowable ranges. Sites are grouped as overall (all sites), impact sites (Sub13 – Sub33) and control sites (Sub40 – Sub48). Note that mercury concentrations are reported as < 0.03 ppm as a result of concentrations being lower than laboratory detection limits. 19

Table 8: Threshold (warning) levels of benthic macrofauna abundance (mean abundance/m<sup>2</sup>) and species richness (number of species) for intertidal and subtidal communities (adapted from Clark et al., 2017). 21

Table 9: Median macrofauna abundance and species richness in intertidal and subtidal impact and control stations as determined in the winter 2021 survey. Values printed in red violate the minimum 80<sup>th</sup> percentile thresholds of the CSMP. 23

Table 10: Mean ( $\pm$  standard error) species richness, abundance (per m<sup>2</sup>) and Shannon-Wiener H' diversity indices of intertidal macrofaunal communities inhabiting the central and northern banks in winter 2021. 24

Table 11: Species that contributed up to 60% to overall similarity within intertidal habitats sampled (central bank, northern bank). 25

Table 12: Mean ( $\pm$  standard error) abundance/m<sup>2</sup>, species richness, and diversity (H' index) of subtidal macrofaunal communities inhabiting the central bank, little lagoon, northern bank and overall. 26

Table 13: Species that contributed up to 60% to overall similarity within subtidal habitats sampled (central bank, little lagoon, northern bank). 28

Table 14: Threshold (warning) levels of chlorophyll-a ( $\mu\text{g}/\text{m}^2$ ) from intertidal and subtidal sediment samples across the four seasons in a year (adapted from Clark et al., 2017). Median levels measured at impact and control sites should not be lower than the 20<sup>th</sup> percentile values and not above the 80<sup>th</sup> percentile values. 31

Table 15: Median chlorophyll-a concentrations ( $\mu\text{g}/\text{m}^2$ ) at intertidal and subtidal impact and control sites as measured in winter 2021. Values are not to drop below the 20% limit and not above the 80% limit. Values printed in red violate these limits. 32

Table 16: Established Guidelines for trace metals in molluscs in different countries including South Africa (maximum acceptable levels) 36

---

Table 17: Threshold (warning) levels of trace metal concentrations for *Perna perna* from control and impact monitoring stations (adapted from Clark et al., 2017). Note that the concentration for mercury (Hg) is given as parts per billion and not parts per million as for other metals. 37

Table 18: Median concentrations of trace metals in *Perna perna* collected throughout the Port of Durban in winter 2021. Medians are given for all sites, impact (site 11, 12, 14, 16, 18-21) and control (site 8, 9, 10, 15, 17, 22). Values printed in red exceed the 90th percentile threshold of the CSMP. National Guideline levels are given for reference to other work. Note that the concentrations for mercury are given as part per million and not parts per billion. \*Cadmium levels below laboratory detection limits. 39

Table 19: Raw laboratory data of trace metal concentrations in brown mussels (*Perna perna*) collected during the winter 2021 survey from 15 channel buoys (control and impact sites) adjacent to the sandbanks in the Port of Durban. Concentrations that exceed the baseline guideline values are printed in red. Values marked with \* were below laboratory detection limits. 40

Table 20: Threshold (warning) levels to be adopted for fish abundance (no. ind./haul), biomass (g/haul) and species richness (no. species/haul) for the impact monitoring stations (adapted from Clark et al., 2017). 42

Table 21: Classification of impact and control sites for fish surveys in the Port of Durban. 43

Table 22: Median values of fish community dynamics at impact and control sites identified during the winter 2021 survey. Values given alongside 80th percentile thresholds stipulated in CSMP baseline study. Values printed in red violate CSMP threshold guidelines. 45

Table 23: Abundance of fish species caught overall (all sites) and at control and impact sites in winter 2021. 46

Table 24: Mass (g) of fish species caught overall (all sites) and at control and impact sites in winter 2021. 47

Table 25: Species richness and Shannon-Weiner H' indices of fish communities surveyed at 14 monitoring stations within the Port of Durban in winter 2021. 48

Table 26: Comparison with threshold levels of bird numbers and species richness recorded for Central Bank and Little Lagoon in the Durban harbour over the sampling period for all bird species. Values in red violate threshold limits. 52

Table 27: Taxonomic composition of common water-associated birds in the Port of Durban over the three-month sampling period (June - August 2021) 55

Table 28: Avifaunal species richness, diversity and evenness per habitat in the Durban harbour over the winter sampling period. No birds were recorded at Island View. 58

## Abbreviations

CSMP	Central Sandbank Mitigation Plan
CTD	Conductivity, temperature, and depth
EA	Environmental Authorisation
ECO	Environmental Control Officer
EIA	Environmental Impact Assessment
EMC	Environmental Monitoring Committee
EMPR	Environmental Management Programme
ERL	Effects Range Limits
ERM	Effects Range Medium
DEA	Department of Environmental Affairs
DEFF	Department of Environment, Forestry & Fisheries
MACE	Marine biodiversity, Aquaculture, Conservation education & Ecophysiology
NOAA	National Oceanic Atmospheric Administration
NTU	Nephelometric Turbidity Units
PPM	Parts Per Million
TOC	Total Organic Carbon
UKZN	University of KwaZulu-Natal
YSI	Yellow Springs Instrument

---

## Executive Summary

GIBB (Pty) Ltd was appointed by Transnet Group Capital, now Transnet National Ports Authority, to conduct environmental monitoring in the Port of Durban as per the requirements of the Central Sandbank Mitigation Plan (Clark et al., 2017). The rationale for the expansion of the Berths is to improve the safety of the Berths and increase the efficiency of the Port (see EIA report for full project description; DEA ref 14/12/16/3/3/2/275). The objective of the monitoring programme is to monitor biotic and abiotic habitats and species count changes measured against the approved baseline which was established per condition 27 of the EA (DEA ref 14/12/16/3/3/2/275) and is contained in Central Sandbank Mitigation Plan (CSMP) (Clark et al., 2017). Monitoring takes place at the sandbank areas and associated infrastructure and is conducted before, during and after the construction and expansion of Berths 203 – 205. As specified in this plan (CSMP), monitoring will be undertaken quarterly during each season (i.e. summer, autumn, winter and spring) for the duration of the sandbank extension period, after which monitoring will be undertaken biannually for the remaining construction period. The quarterly assessment exercise includes monitoring of water and sediment characteristics, benthic microalgae, benthic macrofaunal communities, biomonitoring of mussels, as well as fish and monthly bird surveys on and adjacent to all sandbank habitats in the Port of Durban. This report provides a summary of information gathered during the winter 2021 survey, which commenced on 1 July 2021 and was the 10<sup>th</sup> sampling event of this programme. Note that construction had not yet commenced at the time of this survey, so all results obtained thus far may be interpreted as building on the baseline data reported in Clark et al. (2017).

The survey was carried out by GIBB Environmental in collaboration with the University of KwaZulu-Natal (UKZN), Marine biodiversity, Aquaculture, Conservation education & Ecophysiology (MACE) Laboratory at the UKZN, Westville Campus. Field samples were collected over the period of 1 – 24 July 2021 by GIBB personnel and students from the UKZN MACE Lab. The following components are assessed as part of this monitoring programme: water quality, sediment characteristics, benthic macrofaunal communities, benthic microalgae, mussel biomonitoring, fish- and bird communities.

### Water Quality

Water quality parameters including temperature, salinity, dissolved oxygen, pH, and turbidity were measured using a YSI (Model: EXO 1 – 18H112179) CTD water quality meter. Measurements were taken at 20 sampling stations distributed along the navigation channels around the intertidal and shallow subtidal sandbank areas in the Port. While no specific limits or thresholds have been set for water quality parameters, the majority of these parameters were similar to the results reported in the baseline study. In winter 2021, water temperatures decreased slightly, as expected from seasonal changes. Salinity levels were greater than those recorded in autumn and were within the ranges reported from the baseline study. Turbidity levels were greater than those recorded in autumn and greater than those described in the baseline study. Dissolved oxygen concentrations and pH levels were similar than those reported in the previous survey.

---

## **Sediment Characteristics**

Sediment samples were collected from 64 stations (39 intertidal and 25 subtidal) on sandbanks in the Port. Intertidal and subtidal sites are further divided into impact and control sites, with impact sites being those that will most likely be affected by the planned developments in the Port. Intertidal and subtidal sites on/around the northern bank are control sites, while those on/around the central bank and little lagoon are impact sites. The impact sites are thus also those that are generally more exposed to shipping activity and therefore more susceptible to anthropogenically-induced changes to natural conditions. Several sediment parameters were assessed, including grain size distribution, Total Organic Carbon (TOC) concentration and trace metal concentrations. According to the ranges defined in the baseline study, sediment grain size must be comprised of mostly sandy sediment and very little proportions of mud. The proportions of sandy sediments were slightly below the minimum threshold, while subtidal grain sizes were within allowable limits. As in previous seasons, the TOC contents of intertidal and subtidal sediments were below the minimum thresholds defined in the baseline study and are indicative of a nutrient-deficient system. In summer, concerning concentrations of mercury were reported but these have remained within allowable levels in autumn and winter. Concentrations of the remaining trace metals were also within allowable levels.

## **Benthic Macrofauna**

Intertidal and subtidal benthic macrofaunal samples were collected from the same stations as sediment samples and results of abundance and species richness are contrasted against season-specific limits defined in the CSMP. Abundance and species richness in intertidal and subtidal zones were, as in previous seasons, below the minimum allowable thresholds stipulated in the baseline study. As an important indicator of overall ecosystem health, these trends represent a noteworthy cause for concern.

## **Benthic Microalgae (Microphytobenthos)**

Microalgae are photosynthetically active micro-organisms that contribute significantly to primary production in sediment and pelagic habitats with important trophic linkages with variety of organisms including macrofauna, fish and birds (Pinckney & Zingmark, 1993). In addition to the functional importance of microalgae in estuarine and marine systems, it is also a useful parameter that can be used to assess ecological health and eutrophication levels of these systems (Stevenson & Pan, 1999). The concentration of chlorophyll-a (chl-a) is used as a proxy for the abundance of microalgae. In this survey (winter 2021), median intertidal and subtidal chl-a concentrations increased from autumn and exceeded the maximum threshold concentrations for this season. This was driven by the elevated chl-a concentrations at the northern bank.

## **Biomonitoring Using Mussels (*Perna perna*)**

Samples of brown mussels (*Perna perna*) were collected from 15 out of 16 designated channel buoys (one buoy did not contain mussels) adjacent to the sandbanks and were analyzed at a SANAS accredited laboratory for trace metal concentrations. Apart from mercury, all trace metals were below the maximum threshold values of the baseline study. This is the sixth consecutive season that mercury concentrations have violated these limits and remains a serious health risk that needs to be addressed by Port management authorities.

---

## Fish

The nearshore fish community in the Port of Durban was sampled using a beach seine net at 14 stations along the margins of the main sandbank areas. Fish communities were, as in previous seasons, dominated by the Bald Glassy (*Ambassis dussumieri*) that also contributed the most to overall biomass. The overall abundance, biomass and species richness of fish communities were below the minimum thresholds for winter and notable declines in all of these measures were observed. There is a possibility that low water temperatures on the day of sampling might have influenced these values, although it is still severely low in comparison to the baseline study's observations.

## Birds

Monthly bird counts are conducted in accordance with methodology used in long-term avian monitoring programmes in the Port. This is also the methodology that informed the CSMP. These surveys are taking place in five main habitats across the Port (central bank/Little lagoon, northern sand banks, mangrove habitat, island view sand bank, open water). Waders and wading birds dominated the various functional groups, and piscivores and invertebrate feeders dominated feeding groups. As in previous seasons, abundance was above the minimum thresholds in all three months, but not species richness. The fact that species richness continues to violate the minimum thresholds indicates that the diversity of avian communities across the Port is demonstrating a consistent decline. While these results cannot be attributed to the construction phase of this project, it is likely that they are attributable to anthropogenic impacts such as plastic pollution that can directly impact the health of marine birds.

## Conclusion

The information presented in this report is part of an ongoing quarterly monitoring programme against which the impacts associated with the proposed development at Pier 2 can be assessed. These assessments will inform the Environmental Control Officer (ECO) and Environmental Monitoring Committee (EMC) who will oversee that the sampled parameters for the Port of Durban are maintained during the construction and operational phases of this development. As construction had not yet commenced, the data gathered thus far can be viewed as additions to an ongoing dataset and will ultimately enable more accurate detection of any potential impacts associated with the planned developments when they occur.

**However, factors that require increased attention and potential mitigation measures from Port management authorities are (1) the consistently high concentrations of mercury in *Perna perna* mussels, (2) decreasing macrofaunal abundance and diversity, (3) decreasing fish abundance and diversity, and (4) decreasing avifaunal diversity.**

There are several factors that may be affecting these ecosystem components. As stated in previous reports, these potential factors include the Umbilo river oil spill of October 2020, storm water runoff and associated pollutants entering the Port and the persistent and ever-increasing plastic pollution in this habitat. These factors cannot be viewed as separate from this monitoring programme as they ultimately affect the integrity of the environment in which surveys are taking place. The impacts of both oil and plastic pollution on marine life are widely acknowledged. The bird roosting habitats in the Port presently consists mainly of plastic debris

---

and inevitably affects the health of these animals. Marine birds often mistake plastic for food and can feed it to their chicks. It is also becoming increasingly difficult for fieldworkers to conduct counts on sandbanks as plastic debris are obscuring birds. Plastic that has broken down to smaller sizes (microplastics) are also fed on by fish, which can ultimately result in death as it is indigestible. While the effects of microplastics on benthic macrofauna are still being investigated, current research suggests that these communities are also most likely negatively affected.

---

# 1 Background

The Port of Durban is located on the east coast of South Africa and occupies the natural area of the Durban Bay, an estuarine embayment fed by the uMbilu, uMhlatuzana and aManzimnyama rivers (Whitfield, 1998). It is one of the few natural harbours on the South African coast and is under the jurisdiction of the Transnet National Ports Authority. Since the development of the Durban Container Terminal in 1977, the Port of Durban has experienced consistently high growth of import and export cargo volumes from around the world. Of the eight commercial Ports in South Africa, the Durban Port is currently South Africa's main general cargo and container Port, handling over 80 million tons of cargo per year (Clark et al., 2016).

Transnet National Ports Authority intends to deepen, widen and lengthen Berths 203 to 205 in the Port of Durban and was granted Environmental Authorisation for the work on 21 January 2015 (EIA and EA reference number 14/12/16/3/3/2/275). The objective is to maximize the efficiency of the Port as well as improve the safety of the Berths. Condition No. 27 of the EA required that baseline monitoring be undertaken for 24 months prior to construction. The EA was appealed and on 9 September 2015 the Minister of Environmental Affairs upheld the EA against the appeal (Appeal Decision Ref LSA 141396) wherein under section 4.2.4 the following was added: *“Amend the EA in line with the eThekweni Municipality’s concerns in respect of 4.3.14 below. To this end, the aforementioned EA is amended to include the following condition: the eThekweni Municipality is to be involved in the baseline monitoring of the sandbank and must ensure that the outcomes of the baseline monitoring inform the central sandbank mitigation plan. The monitoring of compliance against the baseline monitoring must solely be the responsibility of the ECO and be independent of the applicant. Construction activities on the central sandbank may only commence upon the successful implementation of the central sandbank mitigation plan”*.

Transnet undertook the 24 months of environmental monitoring within the Durban Bay, and as part of the Environmental Management Programme (EMPR) produced a plan specifically dealing with marine works on and around the Centre Sand Bank (the Central Sandbank Mitigation Plan – CSMP; Clark et al., (2017)).

In line with Clause 4.2.4 of the Appeal Decision, the “implementation” of the CSMP was clarified with then DEA (now the Department of Environment, Forestry and Fisheries; DEFF) to mean:

- 1) the completion and publication of the plan
- 2) the submission to the Authorities and approval of the plan; and
- 3) the issuing of the plan to the contractors prior to commencement of construction activities.

The Department was in agreement with this interpretation, and hence construction could commence.

Through an open tender process GIBB was appointed as the ECO on the project and in light of EA condition 27 and Appeal Decision condition 4.2.4, initiated this monitoring programme



---

for all the sandbank areas in the Port during the proposed construction of the Berths. The intention is to thereby assess the impacts of any disturbances associated with this development such as loss of supratidal, intertidal and subtidal habitats within the Port. Monitoring activities include quarterly surveys of water quality, sediment characteristics, benthic microalgae, benthic macrofauna, mussel biomonitoring, fish and a monthly bird survey on and adjacent to all sandbank habitats during the sandbank works, and thereafter bi-annually for the remainder of the construction period.

The development of the Port of Durban, together with the growth and urbanisation of the surrounding city of Durban has resulted in significant changes in the natural functioning of this estuary. However, Durban Bay is still considered an estuary with high national conservation importance, with an overall importance score of 92% (Turpie & Clark, 2007). It is ranked as the 10th most important estuary in South Africa (Turpie & Clark, 2007). In order to secure the benefits provided by this system in terms of ecosystem goods and services, it has been recommended that at least partial protection be assigned to the estuary. This has become an objective of the Bay of Natal Estuarine Management Plan (ERM/MER, 2012) to protect and enhance estuarine habitats, which are characteristic of the original Bay; and to explore opportunities for rehabilitating/improving and expanding existing soft habitat. It is important to conserve and manage the remaining mangroves, mudflats and sandbanks of Durban Harbour in order to retain the valuable ecological functions that they offer to the entire system.

The intertidal sandbank habitats in the Port are important to its ecological functioning (Newman et al., 2008; Weerts, 2010). They have significant ecological importance as they contribute to the various ecosystem goods and services provided by the Port. Intertidal sandbanks in the Port become exposed at low tide and play an important role in the recycling of terrestrial and marine derived nutrients and organic matter (Deborde et al., 2008). The sandbank habitats are important from a conservation perspective as they harbour a diverse community of invertebrate fauna. They are also important feeding areas for fish and birds and thus help maintain biodiversity in the Port (Allan et al., 1999). The Bay of Natal Estuary Management Plan (MER/ERM, 2012) has accordingly identified the sandbank habitat for conservation.

A number of studies have been completed in recent years focusing on the estuarine biota of the Port of Durban (Allan et al., 1999; Pillay, 2002; Blackler et al., 2004; Forbes & Demetriades, 2006; Angel & Clark, 2008; Newman et al., 2008; Weerts, 2010; MER/ERM, 2012; Clark et al., 2016). Key sandbank habitats in the Port include the Centre Bank, Little Lagoon, Northern Banks and the Mangroves. This document details sampling procedures and information gathered from the winter 2021 environmental monitoring survey.

## **2 Project Location and Baseline Environment**

### **2.1 Port of Durban**

The Port of Durban is considered an estuarine embayment with three perennial rivers that feed into it. The harbour has approximately 57 storm water outfalls, which feed the bay with surface runoff. The total size of the Durban Harbour catchment is approximately 242 km<sup>2</sup>. The surface

area of the Durban harbour bay is approximately 8 km<sup>2</sup>, with a shoreline perimeter of approximately 27 km (Day & Morgan, 1956; Guastella, 1994; Hay et al., 1995).

## 2.2 Geology

The Durban Harbour is considered a bay, which serves as a major receiver of run-off water from canals and surrounding rivers. The Port of Durban is underlain by Cretaceous, aged siltstone rocks of the St Lucia Formation, Zululand Group, that have weathered to varying degrees. Residual soils have been derived from the *in situ* weathering of the bedrock. Younger unconsolidated alluvial and estuarine sediments referred to as the Harbour Beds unconformably overlie the Cretaceous-aged strata in the Port of Durban (Hindmarch et al., 2008; Cawthra et al., 2012).

## 2.3 Site Location

The study site (**Figure 1**) is located within the Port of Durban (29°52'17.87"S; 31° 1'9.08"E). The Port occupies the natural expanse of the full Durban Bay, spanning an area of 1850ha (18.5km<sup>2</sup>) (Transnet National Ports Authority, 2017). The city centre is located approximately 5km north of the Port of Durban, with the residential and industrial area of the Bluff located 10 km south. The Port is one of the few in the world with such a close proximity to the central city.



**Figure 1:** Ariel view of intertidal sandbank habitats (Northern Bank, Centre Bank, Little Lagoon, Island View, Bayhead Mangroves) in the Port of Durban (Google Earth, 2015).

---

## 2.4 Aspects of environmental monitoring

The environmental monitoring presented in this report include monitoring of physico-chemical variables and assemblages of fauna and flora in the Port of Durban.

Physico-chemical variables include:

- i. Salinity
- ii. Temperature
- iii. Dissolved Oxygen
- iv. pH
- v. Turbidity
- vi. Sediment grain size distribution
- vii. Sediment organic carbon content
- viii. Trace metal concentrations in sediment and mussels (Al, As, Cd, Co, Cr, Cu, Fe, Mn, Ni, Pb, Zn, Hg)

Fauna and flora assemblages include:

- i. Benthic microalgae (microphytobenthos)
- ii. Benthic macrofauna
- iii. Fish
- iv. Birds

Detailed methodologies for each of these components are presented in the subsections below. A quarterly (once every 3 months) sampling schedule is being followed for water and sediment quality, benthic microalgae, benthic macrofauna and fish surveys, while a monthly schedule is being followed for bird surveys. Dates of the quarterly surveys were selected to correspond with the middle of each season and are centred around one of the spring tide episodes in each period.

Surveys have been completed in the following months:

- October 2018 (Spring I). Completed.
- January 2019 (Summer I) – This sample run did not occur as Transnet put the project on hold
- May 2019 (Autumn I) – This sample run did not occur as Transnet put the project on hold
- August 2019 (Winter I) – Completed.
- October 2019 (Spring II) – Completed.
- January 2020 (Summer II) – Completed.
- June 2020 (Autumn II) – Completed.
- July 2020 (Winter II) – Completed.
- October 2020 (Spring III) – Completed.
- January 2021 (Summer III) – Completed.
- April 2021 (Autumn III) – Completed.
- July 2021 (Winter III) – Completed. Presented in this report.

---

## 3 Water Quality

### 3.1 Introduction

Estuaries are dynamic water bodies, influenced both by the marine environment and freshwater from catchments. The conditions within estuaries vary both seasonally and diurnally (a result of tide, temperature and sunlight). Estuaries are extremely productive ecosystems due to the combination of high nutrient river water with a shallow, sheltered habitat. Parameters typically measured include salinity, temperature, light intensity, dissolved oxygen, nutrients and chlorophyll-a. These parameters help monitor natural phenomena such as stratification and tidal flushing, as well as impacts caused by human activities and from the catchment. Estuaries are important for their role as nurseries for juvenile marine fish and invertebrates that inhabit these protected and nutrient-rich areas during their developmental stages (Beck et al. 2001). However, water quality in the Durban Bay has become severely degraded because of poor catchment management upstream including erosion, pollution, water abstraction, and harbour development (Durban Port). This presents concern for the biota within the systems.

Estuaries are fragile and sensitive ecosystems that are easily affected by changes in land and developments. Monitoring of water quality is important for the identification of pollution events and for identifying areas of chronic contamination. An example is the introduction of organic waste that can result in deoxygenation, eutrophication and/or the formation of algal blooms with potential adverse impacts on biodiversity (Varadharajan et al., 2013). In addition, Port development activities such as dredging can increase turbidity and total suspended solids in water, reducing light penetration and hinder photosynthetic activities in the water column. This invariably affects the ecosystem functioning of higher organisms that directly depend on primary producers (Brzeski & Newkirk, 1997). Basic water quality of the Port is assessed as part of ongoing monitoring for this project. Water quality parameters that are considered include temperature, salinity, dissolved oxygen content, pH and turbidity.

#### 3.1.1 Temperature

Water temperature is closely associated with many of the chemical and biological processes occurring in estuaries (Snow & Taljaard, 2007). Concentrations and saturation levels of dissolved oxygen in the water is directly influenced by temperature. As water temperature increases, the solubility of oxygen decreases. Therefore, cold water generally holds more oxygen than warmer waters (Madeira et al., 2013). Temperature also influences other biological processes such as photosynthesis, animal reproductive cycles, -migrations, -metabolic rates and susceptibility to parasites and disease (Caffrey, 2003; Pillay & Perissinotto, 2008). Water temperature in estuaries can vary with depth, anthropogenic activities, season, atmospheric conditions and tidal phases. Thermal stratification (i.e. water layers with strong temperature differences) can occur when the mixing of water layers is inadequate. If such conditions persist, it could result in hypoxic / anoxic conditions in bottom waters with potential negative impacts on benthic biota (Chadwick et al., 1996).

---

### **3.1.2 Salinity**

Salinity is a measure of the amount of dissolved salts in water. In estuarine environments, there are significant fluctuations in salinity levels because of the input of freshwater from river inflow or anthropogenic discharge via storm water canals, which can have a direct influence on the type and distribution of plants and animals in an estuary. Salinity levels also fluctuate with season (Kamer & Fong, 2000; Pillay & Perissinotto, 2008). During the rainy season, greater amounts of freshwater enter estuaries thereby lowering salinity levels. In contrast, a dry season could result in increased salinities due to less freshwater coming into the system, and possibly less variation in salinity levels. Just like temperature, salinity stratification could also be a problem in an estuarine environment causing hypoxia and anoxia in benthic habitats.

### **3.1.3 Dissolved Oxygen**

Dissolved oxygen concentration refers to the amount of oxygen present in water and is vital for the survival of most aquatic life. Dissolved oxygen plays an important role in the various chemical and biological processes occurring in estuaries (Reddi et al., 1993). Season, temperature, salinity and plant photosynthetic activity all influence dissolved oxygen levels in an estuary (Attrill et al., 1999). Sufficient levels of dissolved oxygen are crucial for the survival of estuarine fauna and thus have an important influence on their distribution and abundance (Pillay & Perissinotto, 2008). The toxicity of trace metal contaminants such as lead, copper, and zinc can increase with a decrease in dissolved oxygen concentrations. Therefore, the monitoring of dissolved oxygen in estuarine environments is important, as it is a good indicator of estuary health.

### **3.1.4 pH**

pH is a measure of alkalinity or acidity of a solution or substance. It is recorded on a scale of 0 to 14; where 0 – 6 is regarded as acidic and 8 – 14 is alkaline. A pH of 7 is commonly referred to as neutral. In estuaries, it is an important water quality parameter, as slight changes in pH can significantly alter the chemistry of the water and influence biogeochemical processes (Doering, 1996). Changes in pH can influence solubility of some metals such as copper and iron, as well as the toxicity of many harmful compounds such as ammonia (Das & Sahu, 2005). The optimal range of pH values for estuarine fauna is approximately between 6.5 and 8.6 (Clark et al., 2016). pH levels lower than 5 or greater than 9 may be fatal for estuarine fauna and this is often an indication of the introduction of industrial or agricultural pollutants to the estuarine system.

### **3.1.5 Turbidity**

Turbidity is an important measure of water quality especially in estuaries where there is usually mixing and inflow of freshwater from connecting rivers and inland canals (Hellweger et al., 2004). Turbidity is a measure of light penetration in the water column and can be defined as the transparency of water in relation to the amount of light scattered by particulates and dissolved substances, recorded in Nephelometric Turbidity Units (NTU). Monitoring turbidity in aquatic environments is important as changes in turbidity may affect availability of light for plant

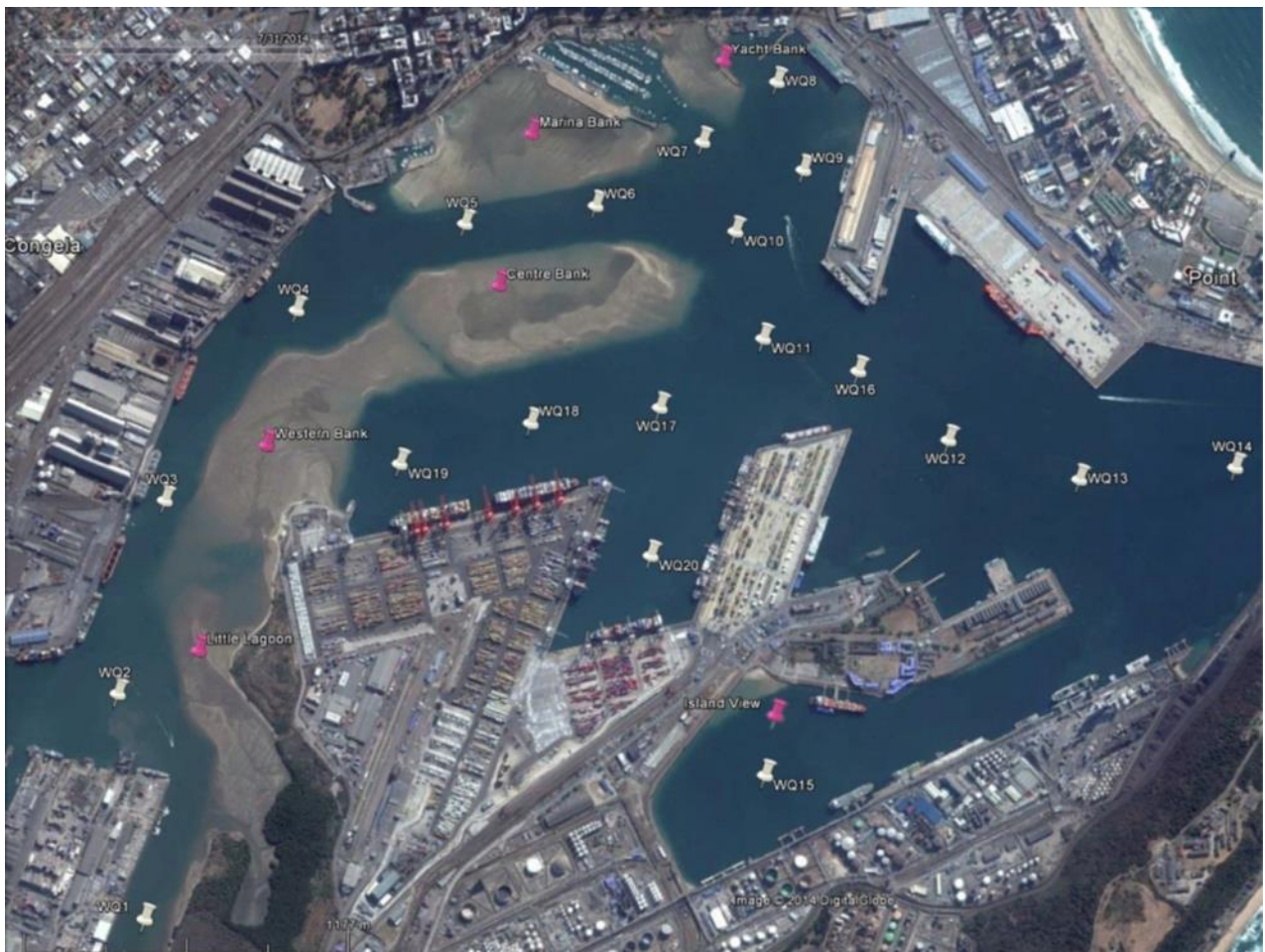


photosynthetic activities, food availability in the system and other ecological processes such as oxygen production (Moore et al., 1997). Elevated turbidity levels can also influence ecological interactions (e.g. predator-prey interaction) through affecting visibility.

### 3.2 Methodology

Water quality parameters including temperature, salinity, dissolved oxygen, pH, and turbidity were measured using a YSI (EXO 1 – 18H112179) CTD water quality meter. Measurements were taken at spring tide (high and low) and neap tide (high and low) at all 20 stations distributed in the navigation channels surrounding the main intertidal and shallow subtidal sandbank areas (**Figure 2**).

The YSI water quality meter was held at the surface for approximately one minute to flush the sensors and was then lowered and retrieved at approximately 1 m/s at each station. Data from all sites were subsequently downloaded and processed according to the manufactures' recommendations. Water quality data from each station were further analysed by deriving appropriate summary statistics to investigate spatial physical-chemical variability in the Port.



**Figure 2:** Water quality monitoring sites (white pins) in the Port of Durban.

---

### 3.3 Results

#### 3.3.1 Temperature

Overall, water temperatures were relatively uniform across the water quality stations in the Port and mean temperatures were lower than those recorded in autumn. Mean surface water temperatures ranged between 18.49 – 20.46°C and mean bottom water temperatures from 19.68– 20.34 °C (**Table 1**), demonstrating no decrease in temperature with depth which suggests that the water column is well-mixed. Overall, temperature readings are similar to those reported in the baseline study (Clark et al., 2017).

#### 3.3.2 Salinity

Salinity values recorded in winter were greater than those recorded in summer and autumn of this year. Specifically, salinity ranges of surface waters ranged from 35.32 – 37.64 PSU and that of deeper waters from 36.7 – 37.69 PSU (**Table 1**). These maximum values were slightly greater than the typical maximum salinity of 36 PSU in marine systems but are still in the range of values reported for this habitat in the baseline study.

#### 3.3.3 Dissolved Oxygen

Dissolved oxygen concentrations in estuaries typically range between 6 – 8 mg/L. Concern is warranted when levels below 2 - 5 mg/L persist over long periods, which can result in physiological stress for most estuarine biota. In winter, mean dissolved oxygen conditions of surface waters were in the range of 7.25 – 9.17 mg/L and 6.21 – 7.37 mg/L in deeper waters (**Table 1**). These concentrations have remained mostly constant throughout the monitoring events completed thus far.

#### 3.3.4 pH

As with other water quality parameters, pH levels varied minimally across all tidal phases throughout the 20 water quality stations in the Port, with means ranging between 6.49 – 7.94 for surface waters and from 7.88 – 7.99 for deeper waters (**Table 1**). pH values recorded at all monitoring stations in the Port appears to remain constant with depth.

#### 3.3.5 Turbidity

Turbidity levels in winter 2021 ranged from 3.69 – 14.14 NTU (Nephelometric Turbidity Units) in surface waters and from 0.3 – 11.5 NTU in deeper water layers (**Table 1**). These values are slightly greater than those reported in the baseline study (i.e. 1 – 7 NTU; Clark et al., 2017).

**Table 1:** Water quality results of surface (< 1 m depth) and bottom layers recorded during the winter 2021 sampling event. All results given as mean  $\pm$  standard deviation.

Station	Temperature (°C)		Salinity (PSU)		Dissolved Oxygen (mg/L)		pH		Turbidity (NTU)	
	Surface	Bottom	Surface	Bottom	Surface	Bottom	Surface	Bottom	Surface	Bottom
WQ01	19.65 $\pm$ 1.57	20.34 $\pm$ 0.47	35.32 $\pm$ 3.82	36.7 $\pm$ 1.32	7.44 $\pm$ 2.24	6.34 $\pm$ 0.86	7.85 $\pm$ 0.08	7.88 $\pm$ 0.04	9.86 $\pm$ 10.38	7.26 $\pm$ 6.99
WQ02	20.46 $\pm$ 0.58	20.32 $\pm$ 0.39	36.3 $\pm$ 1.28	37.37 $\pm$ 0.27	7.7 $\pm$ 1.18	6.21 $\pm$ 0.64	7.94 $\pm$ 0.06	7.95 $\pm$ 0.02	6.05 $\pm$ 3.95	6.15 $\pm$ 11.59
WQ03	19.87 $\pm$ 1.43	20.09 $\pm$ 0.41	35.62 $\pm$ 2.22	37.27 $\pm$ 0.74	7.39 $\pm$ 1.48	6.51 $\pm$ 0.62	7.90 $\pm$ 0.12	7.95 $\pm$ 0.02	6.97 $\pm$ 10.51	2.36 $\pm$ 5.55
WQ04	19.80 $\pm$ 1.3	20.03 $\pm$ 0.47	35.58 $\pm$ 2.95	37.38 $\pm$ 0.35	7.27 $\pm$ 1.58	6.64 $\pm$ 0.65	7.88 $\pm$ 0.28	7.97 $\pm$ 0.01	6.015 $\pm$ 5.92	3.69 $\pm$ 4.15
WQ05	19.7 $\pm$ 1.35	20.05 $\pm$ 0.45	35.69 $\pm$ 4.23	37.34 $\pm$ 0.46	7.28 $\pm$ 1.71	6.58 $\pm$ 0.62	7.93 $\pm$ 0.1	7.95 $\pm$ 0.03	5.84 $\pm$ 11.06	9.36 $\pm$ 20.91
WQ06	20 $\pm$ 0.96	20 $\pm$ 0.53	36.73 $\pm$ 1.52	37.26 $\pm$ 0.42	7.83 $\pm$ 1.03	6.61 $\pm$ 0.59	7.94 $\pm$ 0.1	7.96 $\pm$ 0.03	5.72 $\pm$ 3.02	2.3 $\pm$ 2.95
WQ07	20.08 $\pm$ 0.93	20.02 $\pm$ 0.58	37.14 $\pm$ 0.84	37.39 $\pm$ 0.32	8.19 $\pm$ 1.09	6.71 $\pm$ 0.65	7.93 $\pm$ 0.09	7.98 $\pm$ 0.01	7.73 $\pm$ 2.82	4.93 $\pm$ 7.37
WQ08	19.05 $\pm$ 1.32	19.93 $\pm$ 0.6	37.12 $\pm$ 0.58	37.28 $\pm$ 0.28	7.68 $\pm$ 1.57	6.61 $\pm$ 0.6	7.91 $\pm$ 0.11	7.97 $\pm$ 0.01	7.94 $\pm$ 5.96	11.5 $\pm$ 16.23
WQ09	19.66 $\pm$ 1.16	19.67 $\pm$ 0.48	37.27 $\pm$ 0.20	37.31 $\pm$ 0.27	7.79 $\pm$ 1.41	6.86 $\pm$ 0.56	7.93 $\pm$ 0.14	7.98 $\pm$ 0.01	4.56 $\pm$ 3.69	4.77 $\pm$ 6.88
WQ10	19.95 $\pm$ 1.42	19.74 $\pm$ 0.49	37.51 $\pm$ 0.06	37.51 $\pm$ 0.11	7.99 $\pm$ 1.61	7.04 $\pm$ 0.6	7.82 $\pm$ 0.31	7.96 $\pm$ 0.00	9.51 $\pm$ 11.65	0.3 $\pm$ 0.11
WQ11	19.54 $\pm$ 0.46	19.93 $\pm$ 0.55	37.54 $\pm$ 0.09	37.49 $\pm$ 0.23	8.91 $\pm$ 0.68	6.81 $\pm$ 0.54	7.85 $\pm$ 0.08	7.98 $\pm$ 0.01	9.21 $\pm$ 2.63	7.73 $\pm$ 3.38
WQ12	19.02 $\pm$ 0.62	19.89 $\pm$ 0.55	37.55 $\pm$ 0.20	37.54 $\pm$ 0.26	9.17 $\pm$ 0.82	6.94 $\pm$ 0.53	7.64 $\pm$ 0.17	7.9 $\pm$ 0.01	9.6 $\pm$ 2.4	2.34 $\pm$ 3.46
WQ13	19.15 $\pm$ 0.96	19.96 $\pm$ 0.57	37.48 $\pm$ 0.25	37.62 $\pm$ 1.02	8.86 $\pm$ 0.85	6.93 $\pm$ 0.6	7.89 $\pm$ 0.21	7.9 $\pm$ 0.02	10.21 $\pm$ 4.64	5.6 $\pm$ 7.56
WQ14	20.06 $\pm$ 1.9	19.92 $\pm$ 0.51	37.64 $\pm$ 0.32	37.69 $\pm$ 0.18	8.90 $\pm$ 0.97	7.2 $\pm$ 0.58	6.49 $\pm$ 1.57	7.97 $\pm$ 0.08	14.14 $\pm$ 13.95	9.25 $\pm$ 13.79
WQ15	18.49 $\pm$ 1.57	19.68 $\pm$ 0.7	37.14 $\pm$ 0.67	37.29 $\pm$ 0.54	8.98 $\pm$ 1.37	7.37 $\pm$ 0.59	7.7 $\pm$ 0.59	7.9 $\pm$ 0.12	12.56 $\pm$ 11.42	3.64 $\pm$ 10.42
WQ16	19.81 $\pm$ 1.15	19.93 $\pm$ 0.53	37.33 $\pm$ 0.33	37.55 $\pm$ 0.17	8.6 $\pm$ 0.97	6.79 $\pm$ 0.61	7.9 $\pm$ 0.1	7.99 $\pm$ 0.02	6.95 $\pm$ 3.06	4.88 $\pm$ 3.55
WQ17	19.67 $\pm$ 0.88	20.07 $\pm$ 0.45	36.05 $\pm$ 1.39	37.41 $\pm$ 0.22	7.81 $\pm$ 1.38	6.71 $\pm$ 0.63	7.87 $\pm$ 0.19	7.97 $\pm$ 0.01	6.93 $\pm$ 3.94	6.67 $\pm$ 3.58
WQ18	19.36 $\pm$ 2.05	20.01 $\pm$ 0.48	37.26 $\pm$ 0.82	37.48 $\pm$ 0.18	7.72 $\pm$ 2	6.67 $\pm$ 0.64	7.85 $\pm$ 0.27	7.96 $\pm$ 0.01	7.93 $\pm$ 4.41	6.6 $\pm$ 6.87
WQ19	20.05 $\pm$ 1.28	20.04 $\pm$ 0.47	37.01 $\pm$ 0.97	37.44 $\pm$ 0.45	7.25 $\pm$ 1.25	6.58 $\pm$ 0.57	7.93 $\pm$ 0.08	7.95 $\pm$ 0.03	3.69 $\pm$ 6.63	2.46 $\pm$ 5.48
WQ20	19.29 $\pm$ 0.66	19.93 $\pm$ 0.48	37.6 $\pm$ 0.04	37.46 $\pm$ 0.36	9 $\pm$ 0.68	6.79 $\pm$ 0.54	7.7 $\pm$ 0.21	7.99 $\pm$ 0.01	9.38 $\pm$ 4.4	5.64 $\pm$ 8.29



---

### **3.4 Conclusion**

Physico-chemical conditions of water in the Port were mostly uniform across all the water stations surveyed and have demonstrated typical seasonal fluctuations throughout the monitoring conducted thus far. In the previous sampling season (summer 2021), salinity levels were lower than the typical ranges reported for marine systems, but these ranges increased to more acceptable levels in autumn 2021. Concentrations of dissolved oxygen, turbidity and pH have remained fairly constant from the previous survey and are in line with the baseline conditions reported in Clark et al. (2017). As Berth construction had not begun, these data can be viewed as a compliment to baseline data used to characterize pre-construction water quality conditions.

---

## 4 Sediments

### 4.1 Introduction

Sediments play an important role in terms of habitat provisioning for aquatic organisms. These organisms, in turn, play a role in the establishment of an ecosystem. Several sediment characteristics are frequently incorporated into long-term monitoring programmes, including grain size composition, concentrations of nutrients such as carbon (measured as Total Organic Carbon) and trace metals. Contaminants present in water can settle in the sediment where they can accumulate over time. As such, sediments can be regarded as a pollutant sink (David, 2006). When these sediments are disturbed (e.g. through dredging), contaminants can become suspended in the water column and become available for uptake by marine biota. In addition, sediment characteristics can also be used to explain the community dynamics of macrofaunal invertebrates (Anthony & Héquette, 2007; Watson et al., 2013).

The distribution of different grain sizes are driven mostly by water movement. Finer sediment tends to accumulate in areas with minimal hydrodynamic disturbance, whereas coarser sediment occurs in areas with greater disturbance. Sediments are classified according to grain size (**Table 2**) and studying the composition of grain sizes can reveal areas that are likely to have elevated contaminant levels. This is because pollutants bind more easily to fine or muddy sediments with larger surface areas (Ellingsen, 2002; Austen & Widdicombe, 2006; Anderson, 2008).

**Table 2:** Sediment grain size characterization.

Descriptive term		Grain size
Gravel		> 2000 µm
Sand	Very coarse	1000 – 2000 µm
	Coarse	500 – 1000 µm
	Medium	250 – 500 µm
	Fine	125 – 250 µm
	Very fine	63 – 125 µm
Mud/silt		< 63 µm

Sediment contamination can stem from various activities including wastewater treatment and disposal, runoff, mining, industrial activity, etc. The pollutants introduced from such activities can pose risks to the health and functioning of aquatic ecosystems (Sponseller et al., 2013; Effendi et al., 2016).

Another sediment parameter that serves as a useful metric of overall sediment quality is the concentration of Total Organic Carbon (TOC). While organic matter represents an important food source for organisms such as benthic macrofauna (Lopez et al., 1989; Snelgrove &

Butman 1994), an excess of such nutrients may result in negative impacts on these communities. The degradation process of these nutrients in the sediments requires oxygen and can produce toxic by-products (e.g. ammonia, sulphide) (Gray et al., 2002). Areas that have elevated levels of TOC are also likely to contain other contaminants. The proportions of TOC in sediments can be classified into three broad system types: oligotrophic, mesotrophic and eutrophic (Forbes & Demetriades, 2003) (**Table 3**). Habitats with high TOC levels are at risk to become oxygen depleted (i.e. eutrophic).

**Table 3:** Total Organic Carbon (TOC) classification levels in sediments (adapted from Forbes & Demetriades, 2003).

<b>% Total Organic Carbon</b>	<b>Class</b>	<b>System Type</b>
<0.5%	Very Low	<b>Oligotrophic</b>
0.5 – 1 %	Low	very low levels of nutrients
1 – 2 %	Moderate	<b>Mesotrophic</b> intermediate level of productivity
2 – 4 %	Medium	<b>Eutrophic</b>
>4%	High	rich in nutrients and minerals, can have excessive algal growth and thus diminished oxygen content to the detriment of other organisms

The final sediment characteristic that is considered as part of this monitoring programme is the concentration of trace metals. Trace metals (Al, As, Cd, Co, Cr, Cu, Fe, Mn, Ni, Pb, Zn, Hg) occur naturally in marine environments and play fundamental physiological roles. They tend to accumulate in fine sediments where they are largely passive (non-threatening) but can become harmful when sediments are disturbed (e.g. through dredging) and subsequently suspended in the water column (Summers et al., 1996; Ellingsen, 2002; Austen & Widdicombe, 2006; Anderson, 2008; CSIR, 2016). Metals may be assimilated by organisms such as filter-feeders and can lead to bioaccumulation in higher order marine organisms which may further result in human health consequences if such organisms are consumed.

In this monitoring programme, sediment trace metal concentrations are contrasted against the South African regulations and the Effects Range Low (ERL) and Effects Range Median (ERM) concentrations specified by the National Oceanographic and Atmospheric Administration (NOAA) (**Table 4**). The ERL values (lower 10<sup>th</sup> percentile) and ERM (median) are derived from the literature and refers to the 10<sup>th</sup> percentile and median concentrations of a particular trace metals at which biological effects of trace metal contamination have been observed.

**Table 4:** South African and NOAA sediment quality guidelines for trace element concentrations. Concentrations are parts per million (ppm) dry weight (mg/kg), ERL = Effects Range Low, ERM = Effects Range Median (DEA, n.d).

Metal	South African guidelines		NOAA Guidelines	
	Special Care Range (ppm)	Prohibited (ppm)	ERL (ppm)	ERM (ppm)
Arsenic (As)	30 – 50	>150	8.2	70
Cadmium (Cd)	1.5 – 10	>10.0	1.2	9.6
Chromium (Cr)	50 – 500	>500	81	370
Copper (Cu)	50 – 500	>500	34	270
Mercury (Hg)	0.5 – 5.0	>5.0	0.15	0.71
Lead (Pb)	100 – 500	>500	46.7	218
Nickel (Ni)	50 – 500	>500	20.9	51.6
Zinc (Zn)	150 – 750	>750	150	410

#### 4.1.1 Sediment Threshold Limits set in the Central Sand Bank Mitigation Plan, 2017

Threshold (warning) levels have been established for grain size (% sand and % mud), Total Organic Carbon concentration (%TOC) and trace metals in sediment samples from the Central Sand Bank Mitigation Plan (Clark et al., 2017) (**Table 5**). If these limits are violated, negative impacts on biota such as invertebrates, fish and birds in the Port can occur.

**Table 5:** Baseline threshold (warning) levels for sediment parameters (%sand, mud %, Total Organic Carbon (TOC) %, and trace metal concentrations) in intertidal and subtidal sediment samples from control and impact monitoring stations across the Port of Durban (adapted from Clark et al., 2017). Values for grain sizes (sand, mud) must fall between 10 – 90<sup>th</sup> percentiles whereas values for trace metals must fall below the 90<sup>th</sup> percentile.

Parameter	Intertidal limits		Subtidal limits	
	10 <sup>th</sup> percentile	90 <sup>th</sup> percentile	10 <sup>th</sup> percentile	90 <sup>th</sup> percentile
Sand (%)	99.6	100	97.4	99.9
Mud (%)	0.0	0.4	0.1	2.6
TOC (%)	1.0	2.0	1.4	6.0
As (ppm)	1.734	4.58	1.734	7.73
Cd (ppm)	n/a	0.58	n/a	1.38
Co (ppm)	n/a	3.93	n/a	7.09
Cr (ppm)	n/a	29.93	n/a	54.59
Cu (ppm)	n/a	16.13	n/a	58.88
Fe (%)	n/a	1.14	n/a	2.19
Mn (ppm)	n/a	214.44	n/a	202.83
Ni (ppm)	n/a	7.38	n/a	15.63
Pb (ppm)	n/a	10.94	n/a	46.70
Zn (ppm)	n/a	25.67	n/a	127.26
Hg (ppm)	n/a	0.10	n/a	0.18

## 4.2 Methodology

### 4.2.1 Sampling locations

Sediment samples were taken at 39 intertidal (**Figure 3**) and 25 subtidal (**Figure 4**) sites distributed across the Port.



**Figure 3:** Intertidal sediment sample sites in the Port of Durban. This includes 21 intertidal stations on the existing central sandbank (yellow pins – Int 10-31), 10 intertidal northern bank stations (yellow pins, Int 40-49), and additional 12 intertidal impact-monitoring stations on the central (black pins, Int 2 - 9). Sites immediately adjacent to the newly established sand bank (red pins, Int 53 - 63) will be sampled once development has commenced. Impact monitoring stations: Int2 – Int31, control monitoring stations: Int40 – Int49. Samples are collected at these sites for sediment characteristics, microalgae and benthic macrofauna. The area highlighted in red is where dredging is to take place.





**Figure 4:** Subtidal sites within the Port of Durban where sediment samples are taken. These sites represent three habitats: the central bank (Subt13 – Subt26), the little lagoon (Subt27 – Subt33), and the northern bank (Subt40 – Subt48). Impact monitoring stations: Subt13-Subt33, control stations: Subt40-Subt48. Samples are collected at these sites for sediment characteristics, microalgae and benthic macrofauna. The area highlighted in red is where dredging is to take place.

---

#### 4.2.2 Sampling protocol

Intertidal sediment samples were taken at low tide when these sandbanks were exposed. Intertidal sediment samples were collected using a hand corer with a diameter of 18 cm. Once inserted into the sediment, the sides around the tube were dug out to insert the separator. The top surface of sediment was then separated from the remaining layers of sediment with these remaining layers then discarded. The top layers were subsequently transferred to sample containers and stored in a cooler box with ice.

Subtidal sediment samples were collected using a stainless steel van Veen grab. This grab was attached to 50 m long rope that was fixed onto a vessel so the grab could be manually lowered onto the seafloor for benthic sediment collection. The grab was locked in place with a safety pin prior to deployment, after which it was hoisted over the side of the vessel and lowered until it reached the seafloor where sediment samples were collected. Field scientists standing on the vessel lowered the grab at a constant speed to the seafloor bottom. Once the grab made contact with the seafloor the safety pin triggered (indicated by slack in the lowering wire), the tension on the rope was slowly increased, causing the lever arms to close the grab and take a sample. On retrieval of the van Veen grab, water trapped above sediment in the grab was drained through a bleeder hole, taking care to lose as little fine-grained material as possible. The samples were collected with a Teflon scoop and placed in sterile cups, sealed and labelled with cross-referencing to sampling sites.

Following collection, all intertidal and subtidal sediment samples were placed in individual containers and kept on ice in the field until delivery to the Talbot and Talbot laboratory (SANAS accredited). Once in the laboratory they were stored frozen (-4°C) until further processing and analysis. From these samples, the grain size distribution, Total Organic Carbon (TOC) content and trace metal concentrations were established.

#### 4.3 Results

The CSMP study stipulates that the median values of the various sediment characteristics (grain size, TOC concentration, trace metal concentration) of impact and control sites must be contrasted against the baseline levels established in the CSMP. The key results of all intertidal and subtidal sediment characteristics in relation to these levels are summarised below (**Table 6, Table 7**).

The majority of intertidal sediment characteristics were within allowable limits, apart from grain size proportions and TOC concentrations (**Table 6**), both of which were below their minimum threshold values.



**Table 6:** Key results of intertidal sediment characteristics (grain size, TOC, trace metal concentrations) in relation to the 90th percentile threshold limits of the CSMP. Values for grain sizes (sand, mud) must fall between 10 – 90th percentiles whereas values for trace metals must fall below the 90<sup>th</sup> percentile. Values printed in red are outside of allowable ranges. Sites are grouped as overall (all sites), impact sites (Int3 – Int 31) and control sites (Int40 – Int49). Note that mercury concentrations are reported as < 0.03 ppm as a result of concentrations being lower than laboratory detection limits.

Characteristic	10th percentile	90th percentile	Median (all sites)	Median (impact sites)	Median (control sites)
Sand (%)	99.6%	100%	98.80%	98.54%	99.10%
Mud (%)	0%	0.4%	0.26%	0.3%	0.32%
Total Organic Carbon (%)	1%	2%	0.3%	0.3%	0.27%
As (ppm)	1.7834	4.58	2.57	2.64	2.16
Cd (ppm)	n/a	0.58	0.33	0.34	0.25
Co (ppm)	n/a	3.93	1.24	1.33	0.97
Cr (ppm)	n/a	29.93	10.01	10.6	8.43
Cu (ppm)	n/a	16.13	5.36	5.49	4.26
Fe (%)	n/a	1.14	0.3	0.31	0.24
Mn (ppm)	n/a	214.44	63.46	77.52	34.05
Ni (ppm)	n/a	7.38	2.68	2.72	2.16
Pb (ppm)	n/a	10.94	7.15	7.53	5.96
Zn (ppm)	n/a	25.67	15.79	16.25	12.88
Hg (ppm)	n/a	0.1	< 0.03	< 0.03	< 0.03

Similarly, in subtidal sediments all parameters apart from TOC were within allowable limits (**Table 7**). TOC concentrations were lower than the threshold minimum stipulated in the baseline study.

**Table 7:** Key results of subtidal sediment characteristics (grain size, TOC, trace metal concentrations) in relation to the 90th percentile threshold limits of the CSMP. Values for grain sizes (sand, mud) must fall between 10 – 90<sup>th</sup> percentiles whereas values for trace metals must fall below the 90<sup>th</sup> percentile. Values printed in red are outside of allowable ranges. Sites are grouped as overall (all sites), impact sites (Sub13 – Sub33) and control sites (Sub40 – Sub48). Note that mercury concentrations are reported as < 0.03 ppm as a result of concentrations being lower than laboratory detection limits.

Characteristic	10th percentile (CSMP)	90th percentile (CSMP)	Median (all sites)	Median (impact sites)	Median (control sites)
Sand (%)	97.4%	99.9%	98.9%	98.7%	99.1%
Mud (%)	0.1%	2.6%	0.26%	0.42%	0.1%
Total Organic Carbon (%)	1.4%	6%	0.75%	0.8%	0.65%
As (ppm)	n/a	7.73	2.84	2.79	2.84
Cd (ppm)	n/a	1.38	0.36	0.37	0.34
Co (ppm)	n/a	7.09	1.47	1.56	1.48
Cr (ppm)	n/a	54.59	10.31	10.39	9.55
Cu (ppm)	n/a	58.88	6.8	7.57	6.24
Fe (%)	n/a	2.19	0.33	0.38	0.33
Mn (ppm)	n/a	202.83	81.9	90.05	79.31
Ni (ppm)	n/a	15.63	2.68	2.81	2.32
Pb (ppm)	n/a	46.7	7.91	8.54	8.02
Zn (ppm)	n/a	127.26	18.19	19.3	15.7
Hg (ppm)	n/a	0.18	< 0.03	< 0.03	< 0.03

#### 4.4 Conclusion

Sediment monitoring is being undertaken at 64 sites in the intertidal (39 sites) and subtidal (25 sites) zones of the sandbanks within the Port of Durban. The majority of sediment characteristics align with what those described in the baseline study and are within acceptable levels.

Importantly, mercury concentrations in intertidal and subtidal sediments have decreased from earlier in the year and were within acceptable levels in winter 2021. However, the TOC content in intertidal and subtidal sediments have remained below the minimum allowable threshold for multiple seasons and indicates that the nutrient-deficiency in sediments is a persistent problem. This could be having effects on organisms such as benthic macrofauna that depend on such nutrients for basic physiological functioning.

---

## 5 Benthic Macrofauna

### 5.1 Introduction

The term benthic macrofauna is used to describe the community of small (> 0.5 mm), mainly invertebrate organisms that live upon or within aquatic sandy and muddy substrata (Hauer & Resh, 2017). They play a significant role in the reworking of sediments and assist with enhancing sediment porosity (Goodnight, 1973). This process helps the exchange of oxygen and nutrients in substrates. Macrofaunal organisms are also an important food source for many marine and estuarine predators (Hauer & Resh, 2017) and are fed on by numerous fish, birds and invertebrates (França et al., 2011). Macrofaunal communities within estuaries often have important ecological linkages with neighbouring habitats as certain species from other areas depend on the health of these habitats for their survival (e.g. trophic subsidy) (Weinstein et al., 2005; França et al., 2011). Therefore, degradation or loss of these habitats and communities can have significant effects on ecologically connected environments.

It is important to monitor biological components of the ecosystem in addition to physico-chemical and eco-toxicological variables, as they provide a direct measure of the state of the ecosystem in space and time (O'Brien et al., 2016). Surveys of benthic macrofaunal communities are often done as part of environmental monitoring programmes and these organisms have come to represent a sort of 'bio-indicator' for the health of an aquatic environment. This is because these organisms are sensitive to changes in the surrounding environment (Pinto et al., 2009) and, because they are relatively non-mobile (Gray et al., 2002), they can provide an indication of localised impacts. If a pollution source is known (e.g. discharges, oil spills, dredging activities, other pollutants), it is possible to examine macrofaunal communities along a distance gradient from the pollution source in order to establish the extent of impact. In addition, because these organisms are short-lived, environmental changes will be reflected rapidly in their community compositions (Warwick, 1993).

Various factors can affect the composition of macrofaunal communities. This includes pollutants and environmental variables such as sediment grain size (Riera et al., 2013), brine discharges (Riera et al., 2012), dredging and trace metal concentrations (Ryu et al., 2011; Fonseca et al., 2020) and other pollutants (e.g. sewage (Riera et al., 2013)). Properties of macrofaunal communities that are often examined include abundance, species richness and diversity. Those that are close to a pollution source generally have lower diversity and the densities of different species may be determined by species-specific tolerances to specific pollutants (Pearson & Rosenberg, 1978).

Organic matter is one of the most important pollutants affecting marine life and it can lead to significant changes in community composition and abundance, particularly in semi-enclosed or closed bays such as the Port of Durban where water circulation is limited. High organic loading often results in eutrophication that can alter macrofaunal community dynamics such as growth rates. Resulting anoxic conditions can further drive complete exclusion of certain species based on specific respiratory requirements (Saunders et al., 2007).

The aim for monitoring benthic macrofaunal communities in the Port of Durban is to assess and monitor potential changes in intertidal and subtidal sandbank macrofaunal communities in response to the planned development activities in the Port. The use of dredging equipment to create an extension of the existing sandbanks will create additional shallow intertidal and subtidal sandbank habitats that will be expected to mimic the lost or existing ones, as has been demonstrated in mudflats (Ray, 2000) and saltmarsh (Streever, 2000) systems.

### 5.1.1 Benthic macrofauna threshold limits set in the Central Sandbank Mitigation Plan, 2017

Threshold (warning) levels have been established for the benthic macrofauna from the Central Sand Bank Mitigation Plan, 2017 (**Table 8**). If levels fall below these limits, recovery of invertebrate populations may be delayed and negative impacts on biota such as fish and birds in the port can be expected.

**Table 8:** Threshold (warning) levels of benthic macrofauna abundance (mean abundance/m<sup>2</sup>) and species richness (number of species) for intertidal and subtidal communities (adapted from Clark et al., 2017).

	Spring	Summer	Autumn	Winter
<b><i>Intertidal invertebrates</i></b>				
Abundance (individuals/m <sup>2</sup> )	162.4	209.6	277.6	204.3
Species richness (no. species)	4.0	5.2	6.4	5.6
<b><i>Subtidal invertebrates</i></b>				
Abundance (individuals/m <sup>2</sup> )	232.0	256.0	232.0	176.0
Species richness (no. species)	7.2	8.0	7.2	6.4

## 5.2 Methodology

### 5.2.1 Sampling protocol

Benthic macrofauna samples were collected from the same 64 monitoring stations as for sediments and benthic microalgae along adjacent sandbanks (intertidal and subtidal samples) in the Port (**Figure 3**). Methods used in the baseline report (Clark et al., 2017) were also applied here. Intertidal samples were collected at spring low tide by inserting an 18 cm diameter corer into the sediment to a depth of 30 cm and transferring the contents to a 0.5 mm mesh cone sieve. At each intertidal sampling site, three core samples were taken and kept separate. Contents of the cores were transferred to mesh cones (0.5 mm<sup>2</sup> mesh size) that were agitated until all possible sediment have been removed. The remaining contents were transferred to honey jars and preserved in 90% ethanol. At each subtidal site, three samples were collected

---

using a Van Veen Grab with a bite of 0.085 m<sup>2</sup>. These samples were processed and preserved in the same way as intertidal samples. Subtidal stations had depths of 3 – 8m and were adjacent to sandbanks.

### **5.2.2 Measures of Diversity**

Various measures can be used to assess spatial and/or temporal variation macrofaunal communities. For the purposes of this report, key results are given per habitat (intertidal habitats: central bank, northern bank; subtidal habitats: central bank, northern bank, little lagoon) as well as an overall estimates of intertidal and subtidal communities. The following measures were determined:

- taxonomic composition of each habitat (intertidal habitats: central bank, northern bank; subtidal habitats: central bank, northern bank, little lagoon),
- abundance,
- species richness,
- Shannon-Wiener diversity indices,
- statistical comparisons of overall community compositions and identification of key species

### **5.2.3 Statistical Analysis**

For intertidal habitats, t-tests were used to determine whether there are statistically significant differences in mean abundance, species richness, and Shannon-Wiener diversity indices of communities inhabiting different areas within the Port. A t-test is a statistical test used to compare the means of two groups (e.g. the mean abundance of intertidal macrofauna in the central bank vs the northern bank). For subtidal habitats, similar comparisons were made but in this case a one-way ANOVA was used as this is the appropriate test to use when comparing more than two groups. In this case comparisons were made among communities of the central bank, little lagoon, and northern bank. Both t-tests and ANOVAs produces p-values, whereby a p-value smaller than 0.05 indicates that there is a statistically significant difference among the means of the groups compared. A p-value larger than 0.05 indicates that the groups compared do not differ significantly. These analyses were conducted in R (R Core Team, 2020).

Multivariate statistics can be used to determine whether there are significant differences in the compositions of communities across space/time. To do this, a PERMANOVA test was used. This test operates based on abundance and the presence/absence data of a species in a given habitat. Comparisons were made between intertidal macrofaunal communities inhabiting the central vs northern bank, and among subtidal communities inhabiting the central bank, little lagoon, and the northern bank. After assessing potential differences in communities inhabiting different areas in the Port, a SIMPER analysis was performed to determine which species distinguished the various communities. The results from this test also provides an indication of species most likely to drive differences (if they exist) among communities. These analyses were carried out through the PRIMER (v6) software.

## 5.3 Results

### 5.3.1 Key results

The CMSP baseline study stipulates minimum limits for parameters such as macrofauna abundance and species richness. These parameters are not to fall below their respective minimums during the construction phase of this project. All parameters in intertidal and subtidal habitats were substantially lower than the minimum thresholds specified for autumn (**Table 9**) and demonstrate further declines from the last sampling event in autumn 2021.

**Table 9:** Median macrofauna abundance and species richness in intertidal and subtidal impact and control stations as determined in the winter 2021 survey. Values printed in red violate the minimum 80%tile thresholds of the CSMP.

	80% limit	Median (overall)	Median (impact)	Median (control)
<b><i>Intertidal</i></b>				
Abundance	204.3	91.7	78.6	150.65
Species richness	5.6	4	4	6
<b><i>Subtidal</i></b>				
Abundance	176	40	46	28
Species richness	6.4	6	6	6

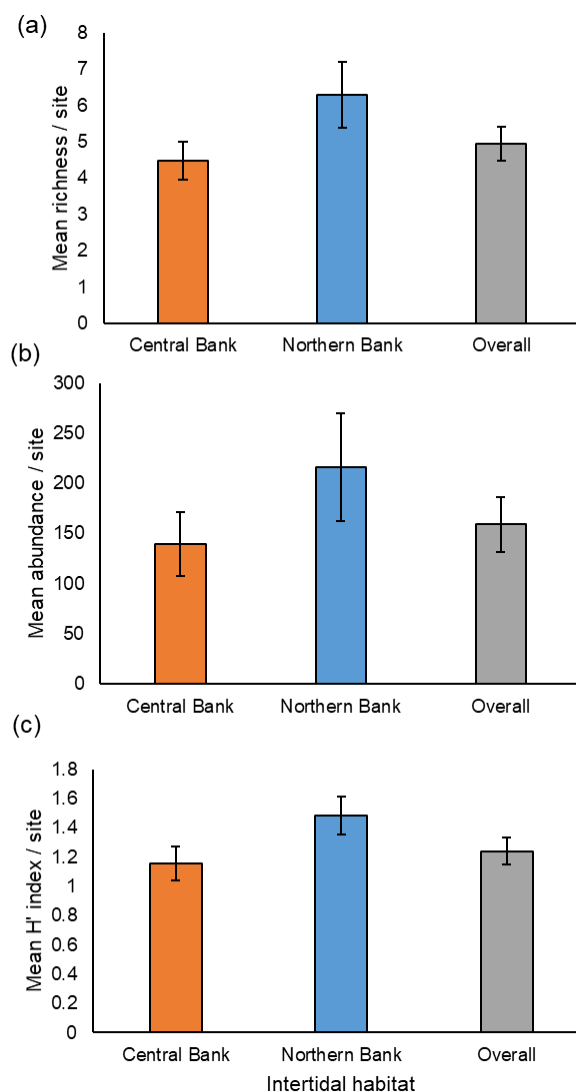
### 5.3.2 Intertidal communities

#### *Abundance, species richness, and diversity*

This winter, a total of 38 intertidal species were recorded, the majority of which were from the central bank ( $n = 34$ ), with fewer on the northern bank ( $n = 25$ ). There were no significant differences in abundance ( $T_{15.902} = -1.233$ ,  $p > 0.05$ ), species richness ( $T_{15.58} = -1.7302$ ,  $p > 0.05$ ) and Shannon-Weiner diversity indices ( $T_{24.49} = -1.8572$ ,  $p > 0.05$ ) between macrofaunal communities on the central and northern banks (**Table 10, Figure 5**).

**Table 10:** Mean ( $\pm$  standard error) species richness, abundance (per m<sup>2</sup>) and Shannon-Wiener H' diversity indices of intertidal macrofaunal communities inhabiting the central and northern banks in winter 2021.

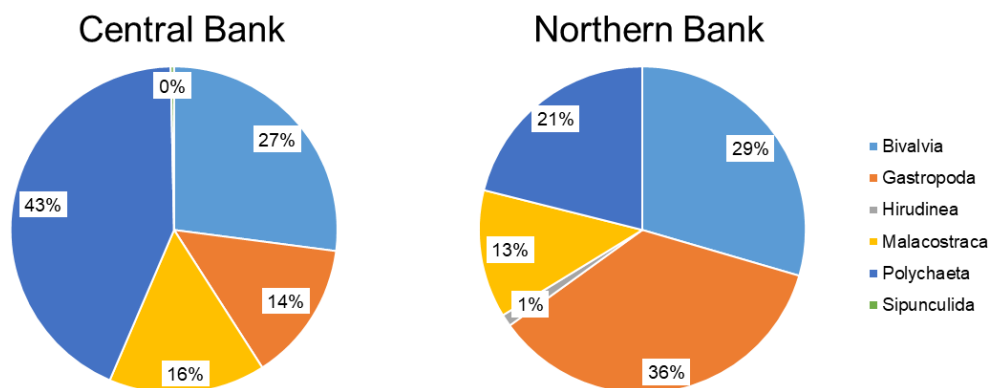
	Central Bank	Northern Bank	Overall
Abundance/m <sup>2</sup>	139.59 ( $\pm$ 31.83)	216.15 ( $\pm$ 53.35)	159.22 ( $\pm$ 27.52)
Species richness	4.48 ( $\pm$ 0.53)	6.3 ( $\pm$ 0.9)	4.95 ( $\pm$ 0.47)
Shannon-Wiener H'	1.16 ( $\pm$ 0.12)	1.48 ( $\pm$ 0.13)	1.24 ( $\pm$ 0.1)



**Figure 5:** Mean ( $\pm$  standard error) (a) species richness, (b) abundance and (c) diversity of intertidal macrofaunal communities of the central and northern sandbanks.

### *Taxonomic composition*

Similar classes were dominant in the two intertidal habitats, although gastropods (snails) were substantially more dominant on the northern bank than the central bank (**Figure 6**), while polychaetes (bristle worms) were nearly twice as abundant on the central bank than the northern bank.



**Figure 6:** Percentage abundance of various macrofaunal classes in communities on the central bank and northern bank.

#### Comparisons among communities

As in previous seasons, intertidal macrofaunal community composition differed significantly between the central and northern banks (PERMANOVA, Pseudo-F = 2.918,  $p = 0.002$ ). This was further confirmed through results from the SIMPER analysis demonstrating that different species dominated each habitat (

**Table 11).**

**Table 11:** Species that contributed up to 60% to overall similarity within intertidal habitats sampled (central bank, northern bank).

Species	Contribution (%)	Cumulative contribution (%)
<b>Central Bank</b>		
<b>(average similarity = 15.11%)</b>		
<i>Orbinia</i> spp.	27.75	27.75
<i>Spiroplax spiralis</i>	17.51	45.26
<i>Aquillaspio sexoculata</i>	12.21	57.47
<i>Orbinia bioreti</i>	5.8	63.27
<b>Northern Bank</b>		
<b>(average similarity = 22.04%)</b>		
<i>Nassarius kraussianus</i>	26.72	26.72
<i>Dosinia hepatica</i>	23.28	50
<i>Leodamas johnstonei</i>	10.79	60.79



---

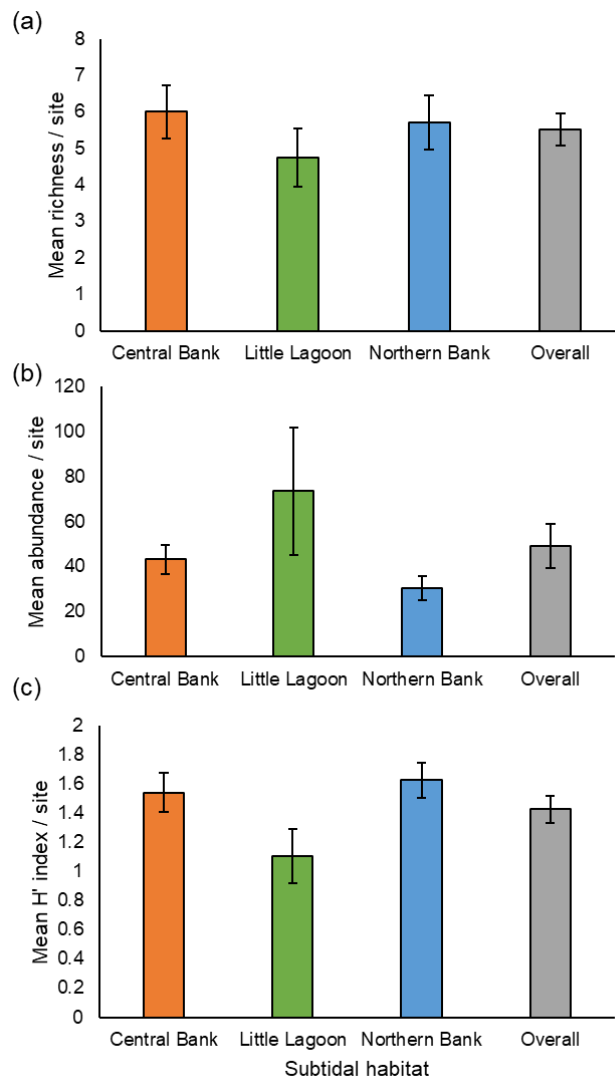
### 5.3.3 Subtidal communities

#### *Abundance, species richness, and diversity*

A total of 27 species were detected from subtidal habitats, the central bank having the most species ( $n = 24$ ) followed by the northern bank ( $n = 16$ ) and then the little lagoon ( $n = 13$ ). There were no significant differences in abundance ( $F_2 = 1.687$ ,  $p > 0.05$ ), species richness ( $F_2 = 0.752$ ,  $p > 0.05$ ) or Shannon Weiner diversity indices ( $F_2 = 3.214$ ,  $p > 0.05$ ) among the three subtidal habitats (**Table 12, Figure 7**).

**Table 12:** Mean ( $\pm$  standard error) abundance/m<sup>2</sup>, species richness, and diversity (H' index) of subtidal macrofaunal communities inhabiting the central bank, little lagoon, northern bank and overall.

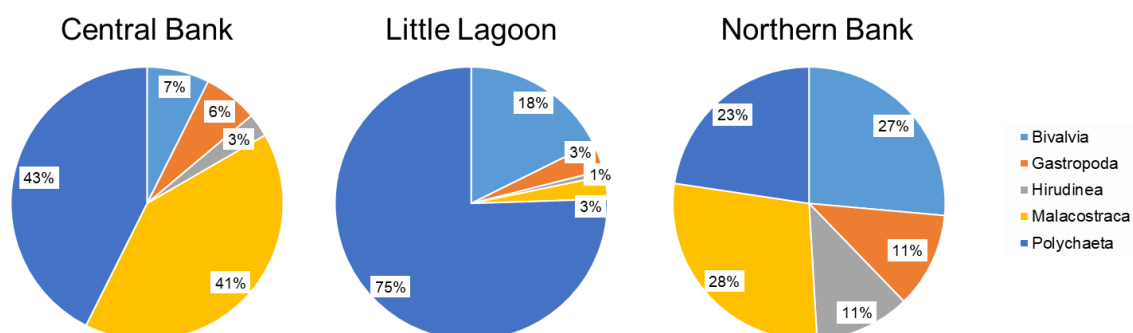
	Central Bank	Little Lagoon	Northern Bank	Overall
Abundance/m <sup>2</sup>	43.2 ( $\pm 6.47$ )	73.5 ( $\pm 28.19$ )	30.29 ( $\pm 5.37$ )	49.28 ( $\pm 9.75$ )
Species richness	6 ( $\pm 0.73$ )	4.75 ( $\pm 0.79$ )	5.71 ( $\pm 0.75$ )	5.52 ( $\pm 0.44$ )
Shannon-Wiener H'	1.54 ( $\pm 0.13$ )	1.11 ( $\pm 0.19$ )	1.63 ( $\pm 0.12$ )	1.43 ( $\pm 0.09$ )



**Figure 7:** Mean ( $\pm$  standard error) (a) abundance, (b) species richness and (c) diversity of subtidal macrofaunal communities of the central bank, northern bank and little lagoon.

#### *Taxonomic composition*

The central and northern banks were dominated by polychaetes, malacostracans and bivalves while in the little lagoon the abundance of polychaetes far outnumbered those of other groups (**Figure 8**). Far fewer malacostracans were recorded in the Little Lagoon than in the other two habitats. Bivalves and gastropods achieved greatest abundances at the Northern Bank.



**Figure 8:** Percentage abundance of subtidal macrofauna classes in communities of the central bank, little lagoon and northern bank.

### Comparisons among communities

As in autumn, there was a significant difference in the composition of subtidal communities (PERMANOVA, Pseudo-F = 2.9677,  $p = 0.001$ ). Pair wise comparisons revealed that all three subtidal habitats differed significantly ( $p < 0.05$ ) from each other. The SIMPER analyses indicated that different species delivered the greatest contributions to respective community similarities, where central bank and little lagoon communities were characterized by the polychaete *Aquilaspio sexoculata*, northern bank communities were characterized by the three-legged crab *Spiroplax spiralis* (**Table 13**).

**Table 13:** Species that contributed up to 60% to overall similarity within subtidal habitats sampled (central bank, little lagoon, northern bank).

Species	Contribution (%)	Cumulative contribution (%)
<b>Central Bank</b>		
(average similarity = 26.97)		
<i>Aquilaspio sexoculata</i>	25.21	25.21
<i>Callichirus kraussi</i>	17.21	42.33
<i>Cirolana fluviatilis</i>	16.84	59.17
<i>Spiroplax spiralis</i>	15.72	74.89
<b>Little Lagoon</b>		
(average similarity = 27.95)		
<i>Aquilaspio sexoculata</i>	52.22	52.22
<i>Dosinia hepatica</i>	9.53	61.75
<b>Northern Bank</b>		
(average similarity = 39.44)		
<i>Spiroplax spiralis</i>	30.23	30.23
<i>Eumarcia paupercula</i>	27.57	57.79
<i>Hirudinea</i>	18.04	75.83

---

## 5.4 Conclusion

As highlighted in previous seasonal report, there are concerning trends regarding macrofaunal biodiversity that are starting to intensify and are not showing signs of improvement. For another season, abundance and species richness and intertidal and subtidal communities did not meet the minimum thresholds stipulated in the CSMP. Although these measures are expected to be lower in winter, they are still too low in comparison to baseline levels. These trends are not related to the construction phase of this project, but this information is still a valuable reflection of the overall health of the Port ecosystem. The overall cause for these trends is most likely pollution, for example from oil spills and the ever-increasing amount of plastic debris in the Port. As emphasized in previous reports, plastic debris that becomes inundated with sand can affect sediment porosity, a habitat feature on which macrofaunal organisms depend (**Figure 9**). Furthermore, as these debris breaks down in the marine environment, they are converted to microplastics that can have further negative effects on the feeding behaviour of macrofaunal organisms.



**Figure 9:** Holes made by common sandprawn *Callichirus kraussi* through a plastic bag. (January 2021)

---

## 6 Benthic Microalgae (Microphytobenthos)

### 6.1 Introduction

Algae is a term used for a diverse group of microorganisms living in the ocean. Microalgae are photosynthetically active microorganisms. Benthic algae, also known as microphytobenthos, refers to microscopic, unicellular photoautotrophs that usually occur in the upper few centimeters of shelf sediments. Below this level, insufficient sunlight hampers their growth. Where the bottom is sandy or muddy the benthic algae cannot attach themselves as they have no roots. Benthic algae deliver major contributions to food webs and biogeochemistry in aquatic ecosystems, energy and cover for many other organisms (Christiansen et al., 2012). In this way, the productivity of the benthic algae in shallow waters directly or indirectly affects the efficiency of the entire marine ecosystem. Benthic microalgae are ubiquitous in aquatic areas where sunlight reaches the sediment surface. Although they are part of the microbial community already discussed, they deserve special attention because they are a very important component of benthic communities in shallow water systems (N-Uptake, 1999). Besides the fact that they contribute significantly to the primary productivity in sediments and pelagic habitats, they also have important trophic linkages with a variety of organisms, including macrofauna, birds and fish (Davis et al., 2015).

Like other single-celled “plant-like” organisms, they use energy gained from sunlight to transform carbon into organic matter via photosynthesis. They generally live in the top few millimeters to centimeters of aquatic sediment but may go deeper in sandy sediments when light availability is very high. In order to grow, benthic microalgae require nitrogen, phosphorus, and other micronutrients in addition to carbon dioxide. Much of the carbon they take up during photosynthesis is released as extracellular polymeric substances (EPS or “slime”) into the sediment (Wolfstein & Stal, 2002). EPS plays an important role in sticking sediment particles together, which may increase sediment stabilization and, thereby, reducing resuspension. In addition, EPS is rapidly metabolized by the bacterial community.

Benthic microorganisms (microalgae and bacteria) are instrumental in controlling the exchange of nutrients, such as nitrogen and phosphorus, across the sediment-water interface (Paerl & Pinckney, 1996). Benthic microalgae in particular, may help to buffer the water column from eutrophication by storing nutrients that would otherwise be used by phytoplankton and bacteria in the water column. Nutrients used by benthic microalgae are derived not only from the water column but also from the sediments themselves as bacteria break down organic matter (Tengberg et al., 2003). Because of their location near the sediment surface, benthic microorganisms play a role in capping the sediments and reducing the release of nutrients to the overlying water column. Nutrients taken up by benthic microalgae and bacteria can be passed up the food chain, especially to meiofauna.

Microalgal communities in the Port of Durban occur on the sandbanks and bottom sediments as microphytobenthos and in the water column as phytoplankton. Microphytobenthos, however, is often suspended in the water column and can be as important as phytoplankton, particularly in turbid estuaries such as in the Port of Durban. Microphytobenthos play a vital

and functional role in estuarine systems. As such, monitoring these communities is an important tool in the overall ecological health assessment of the Port ecosystem.

### 6.1.1 Benthic Microalgae Threshold Limits set in the Central Sandbank Mitigation Plan, 2017

Threshold (warning) levels have been established for benthic microalgae in the Central Sand Bank Mitigation Plan (CSMP) (Clark et al., 2017) (**Table 14**). According to the CSMP, the biomass of benthic microalgae, measured as the concentration of chlorophyll-a ( $\mu\text{g}/\text{m}^2$ ) should not drop below the 20<sup>th</sup> percentile value of the median from baseline data, and should not rise above the 80<sup>th</sup> percentile of these data (**Table 14**). Comparisons are to be made separately for impact and control sites in intertidal and subtidal habitats. Intertidal impact sites are those located on the central bank (Int2 – Int31) and control sites are those on the northern bank (Int40 – Int49). Similarly, subtidal impact sites are those around the central bank and little lagoon (Sub13 – Sub33), while control sites are those around the northern bank (Sub40 – Sub48).

**Table 14:** Threshold (warning) levels of chlorophyll-a ( $\mu\text{g}/\text{m}^2$ ) from intertidal and subtidal sediment samples across the four seasons in a year (adapted from Clark et al., 2017). Median levels measured at impact and control sites should not be lower than the 20<sup>th</sup> percentile values and not above the 80<sup>th</sup> percentile values.

	Spring	Summer	Autumn	Winter
<b><i>Intertidal</i></b>				
20%	6.3	7.7	1.4	0.6
Median	22.6	17.7	12.3	2.5
80%	47.7	66.4	43.7	21.6
<b><i>Subtidal</i></b>				
20%	1.5	6.6	1.6	1.1
Median	3.7	11.5	6.5	3.0
80%	8.2	21.2	26.8	8.3

## 6.2 Methodology

### 6.2.1 Sampling protocol

Benthic microalgae samples were collected from each of the 64 sediment monitoring sites and analyzed in accordance with methods prescribed by Pinckney and Zingmark (1993). Samples were collected by inserting a 20 mm vial directly into the sandbank sediment for intertidal samples, or into the top layer of sediment collected by an Ekman grab sampler for subtidal samples. From both habitats, a sample core of 40 mm in length was extracted and immediately sealed off with a plastic lid. Samples were immediately placed on ice in a dark container and transported to the laboratory for chlorophyll-a analysis.

### 6.2.2 Laboratory protocol

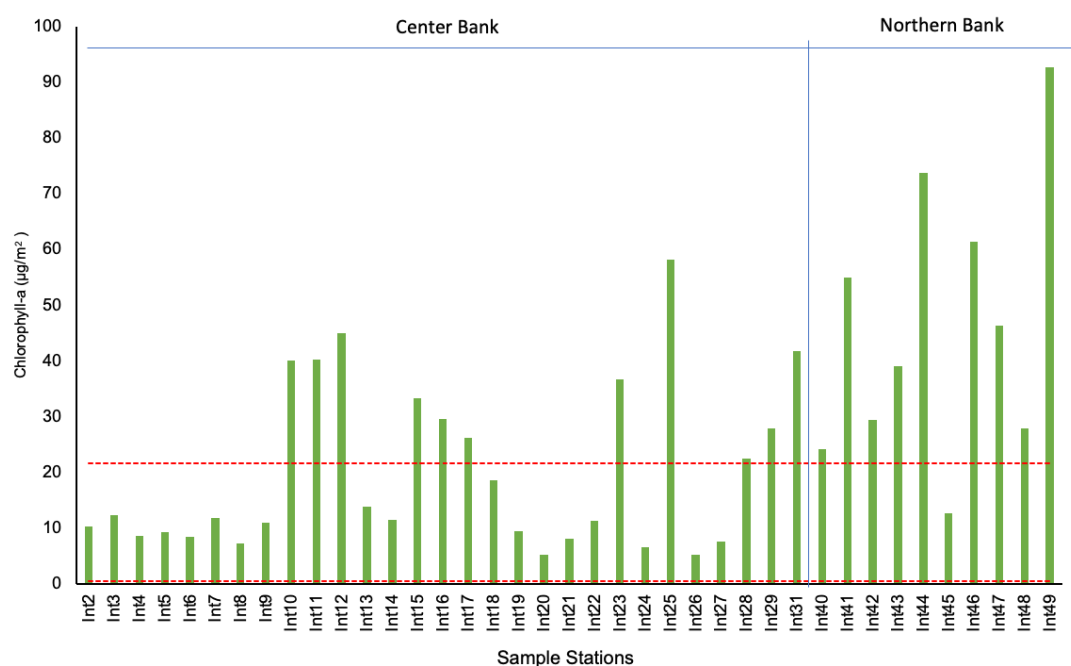
In the laboratory, the biomass of microalgae in the sediment samples was estimated as total chlorophyll (Chl-a) according to the methods of Whitney & Darley (1979), Dandonneau & Neveux (2002) and Seuront & Leterme (2006). Chlorophyll was extracted from the sediment samples through addition of 8 – 10 mL of 90% acetone. This mixture was then centrifuged for approximately 5 minutes at 8000rpm. For each sample, 1mL of the supernatant containing the chlorophyll was pipetted into microfuge tubes and analyzed using a Trilogy Turner Fluorometer™, which yielded results of chlorophyll-a concentrations in  $\mu\text{g/L}^{-1}$  at each site. 1 - 2 drops of 1 M HCl was used to eliminate interference from degraded pigments that are common in sediment samples (Cahoon & Cooke, 1992).

### 6.3 Results

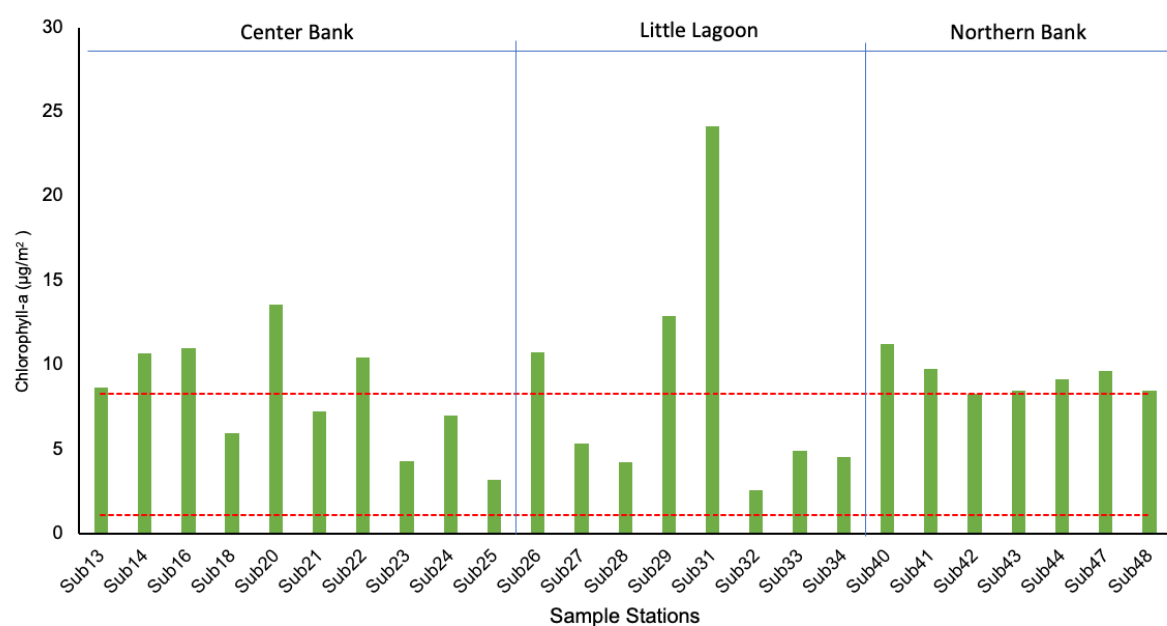
The median chl-a concentrations of control and impact sites in intertidal and subtidal habitats were above the allowable maximum values in intertidal and subtidal habitats (**Table 15**). This was driven by elevated concentrations at control sites (i.e. northern bank) in intertidal and subtidal habitats. As in previous seasons, subtidal chl-a concentrations were lower than those of intertidal habitats. The greatest intertidal chlorophyll-a concentration was  $92.66 \mu\text{g/m}^2$  (Int49) at the Northern Bank (**Figure 10**). The greatest subtidal chlorophyll-a concentration of  $24.17 \mu\text{g/m}^2$  was recorded at station Sub31 in the Little Lagoon area (**Figure 11**).

**Table 15:** Median chlorophyll-a concentrations ( $\mu\text{g/m}^2$ ) at intertidal and subtidal impact and control sites as measured in winter 2021. Values are not to drop below the 20% limit and not above the 80% limit. Values printed in red violate these limits.

	20% limit	80% limit	Median (overall)	Median (impact)	Median (control)
<b><i>Intertidal</i></b>					
Chl-a ( $\mu\text{g/m}^2$ )	0.6	21.6	22.51	11.79	42.73
<b><i>Subtidal</i></b>					
Chl-a ( $\mu\text{g/m}^2$ )	1.1	8.3	8.5	7.11	9.14



**Figure 10:** Chlorophyll-a concentrations ( $\mu\text{g}/\text{m}^2$ ) in 39 intertidal monitoring stations in the Port of Durban, winter 2021. Impact sites = Int2 – Int31 (central bank), control sites = Int40 – Int49 (northern bank). Red dotted lines: 20%tile ( $0.6 \mu\text{g}/\text{m}^2$ ) and 80%tile ( $21.6 \mu\text{g}/\text{m}^2$ ) intertidal winter threshold limits stipulated in the CSMP.



**Figure 11:** Chlorophyll-a concentrations ( $\mu\text{g}/\text{m}^2$ ) in 25 subtidal monitoring stations in the Port of Durban, winter 2021. Impact sites = Sub13 – Sub34, control sites = Sub40 – Sub48. Red dotted lines: 20%tile ( $1.1 \mu\text{g}/\text{m}^2$ ) and 80%tile ( $8.3 \mu\text{g}/\text{m}^2$ ) subtidal winter threshold limits stipulated in the CSMP.



---

## 6.4 Conclusion

Chlorophyll-a and light availability are major determinants of benthic primary production (MacIntyre et al. 1996). Microphytobenthos may also contribute significantly to production in the water column when sediment is re-suspended by currents, upwelling or other forms of disturbance. These algae and cyanobacteria are an important food source for deposit-feeding and suspension-feeding macrofauna. An abundance of microphytobenthos can result in a greater diversity of species higher up the food chain and plays a significant role in system productivity and trophic dynamics.

The chlorophyll-a levels recorded in winter 2021 were greater than those recorded in autumn. The median chl-a levels at impact and control sites were above the maximum baseline threshold stipulated in the baseline study (Clark et al., 2017). These trends need to be monitored closely in upcoming seasons to determine if they are persistent.

---

## 7 Biomonitoring using Mussels (*Perna perna*)

### 7.1 Introduction

Trace or heavy metals are persistent pollutants in aquatic ecosystems. They are all naturally occurring chemical elements, some of which (e.g. copper and zinc) are required by organisms in considerable quantities (Phillips, 1980). Aquatic organisms will normally accumulate essential trace metals that occur naturally in water. However, these metals can be toxic at greater concentrations (Rainbow, 1995). Anthropogenic activities and other naturally occurring geochemical processes may significantly increase the rates of mobilization of trace metals from the earth's crusts, which can lead to increases in their bioavailability in coastal waters through runoffs and wastewater discharge (Phillips, 1995).

Even though monitoring of heavy metal concentrations in sediments is more preferable when resolving analytical and temporal variability problems due to their accumulation in aquatic sediments, this method does not provide accurate information regarding their bioavailability to organisms inhabiting these environments. Measuring metal concentrations in the tissues of aquatic organisms appears to be the most suitable method for assessing eco-toxicity as the metals are frequently accumulated in high (easily measurable) concentrations and reflect a time-integrated measure of bioavailable metal levels (Rainbow, 1995).

Long-term pollution impacts on marine environments is increasingly being monitored through the assessment of contaminants that build up in marine biota. Filter feeding organisms such as mussels have been used successfully as bio-indicator organisms in environmental monitoring programs throughout the world (Kljaković-Gašpić et al., 2006, 2010). They are suitable indicators because they are abundant, widely distributed, sessile, tolerant of salinity changes, stress-resistant and can accumulate a wide range of contaminants (Phillips & Rainbow, 1993; Desideri et al., 2009; Kljaković-Gašpić et al., 2010). In addition, mussels can accumulate trace metals, hydrocarbons and pesticides in their flesh (Rainbow et al., 2006). As such, assessments of contaminant levels in mussel flesh can be used to detect pollution levels that may go unnoticed between pollution events or that may occur at chronically low levels that can be difficult to measure in the water column. Monitoring contaminant levels in mussels can therefore provide a reliable indication of water quality and spatial/temporal changes in bioavailable contaminant levels in the water column. For this purpose, the trace metal concentrations in mussels (*Perna perna*) throughout the Port of Durban are assessed.

#### 7.1.1 Trace metal concentration guidelines for molluscs

Several guidelines are available regarding the acceptable concentrations of trace metals in animals such as molluscs (e.g. mussels) (**Table 16**). In addition, the CSMP (Clark et al. 2017) also provides local maximum thresholds specifically for *Perna perna* that should be assessed as part of this monitoring programme (**Table 17**). If levels exceed limits specified below, risks to health of people consuming shellfish and fish from the Port are likely to be elevated and negative impacts on biota such as invertebrates, fish and birds in the Port are likely.

**Table 16:** Established Guidelines for trace metals in molluscs in different countries including South Africa (maximum acceptable levels)

Country	Cu (ppm)	Pb (ppm)	Zn (ppm)	As (ppm)	Cd (ppm)	Hg (ppm)
South Africa <sup>1</sup> .	-	0.5	-	3.0	3.0	0.5
Canada <sup>2</sup>	70	2.5	150.0	1.0	2.0	-
Australia & NZ <sup>3</sup>	-	2.0	-	-	2.0	0.5
European Union <sup>4</sup>	-	1.5	-	-	1.0	0.5
Japan <sup>5</sup>	-	10.0	-	-	2.0	0.2
Switzerland <sup>2</sup>	-	1.0	-	-	0.6	0.5
Russia <sup>6</sup>	-	10.0	-	-	2.0	-
South Korea <sup>2</sup>	-	0.3	-	-	-	-
United States <sup>7, 8</sup>	-	1.7	-	-	4.0	-
China <sup>9</sup>	-	-	-	-	2.0	-
Brazil <sup>10</sup>	-	-	-	-	-	0-5
Israel <sup>10</sup>	-	-	-	-	-	1.0

1. Regulation R.500 (2004) published under the Foodstuffs, Cosmetics and Disinfectants Act, 1972 (Act 54 of 1972)
2. Fish Products Standard Method Manual, Fisheries & Oceans, Canada (1995)
3. Food Standard Australia and New Zealand (website)
4. Commission Regulation (EC) No. 221/2002
5. Specifications and Standards for Foods. Food Additives, etc. Under the Food Sanitation Law JETRO (Dec 1999)
6. Food Journal of Thailand. National Food Institute (2002)
7. FDA Guidance Documents
8. Compliance Policy Guide 540.600
9. Food and Agricultural Import Regulations and Standards.
10. Fish Products Inspection Manual, Fisheries and Oceans, Canada, Chapter 10, Amend. No. 5 BR-1, 1995.

In addition to the national guidelines, local guidelines for trace metal concentrations in *P. perna* have also been established as part of the baseline study (**Table 17**).

**Table 17:** Threshold (warning) levels of trace metal concentrations for *Perna perna* from control and impact monitoring stations (adapted from Clark et al., 2017). Note that the concentration for mercury (Hg) is given as parts per billion and not parts per million as for other metals.

Trace metal	90 <sup>th</sup> Percentile
As (ppm)	3.61
Cd (ppm)	1.08
Co (ppm)	0.95
Cr (ppm)	8.44
Cu (ppm)	23.18
Fe (ppm)	800.11
Mn (ppm)	134.75
Ni (ppm)	5.60
Pb (ppm)	8.95
Zn (ppm)	223.88
Hg (ppb)	293.06

## 7.2 Methodology

As per the CSMP, mussels (*Perna perna*) must be collected from 16 channel buoys adjacent to the sandbanks in the Port (**Figure 12**). Sites 11, 12, 14, 16, 18 – 21 have been designated as impact sites, and sites 8 – 10, 13, 15, 17, 22 and 23 are control sites. In this survey, mussels could be collected from 15 channel buoys (one buoy did not contain any mussels). Mussels collected were placed on ice in plastic containers and transported to a SANAS accredited analytical laboratory for trace metal (Al, As, Cd, Co, Cr, Cu, Fe, Mn, Ni, Pb, Zn, Hg) analyses. Trace metal concentrations are contrasted against the 90th percentile maximum threshold levels prescribed in the CSMP (Clark et al., 2017) and against the maximum legal limits prescribed for each contaminant in shellfish for human consumption in South Africa, as stipulated by the Regulation R.500 (2004) published under the Foodstuffs, Cosmetics and Disinfectants Act, 1972 (Act 54 of 1972). Where threshold values have not been specified in national legislation, those adopted by other countries were employed.



**Figure 12:** Locations of channel buoys for brown mussel (*Perna perna*) biomonitoring in the Port of Durban. Impact sites: 11, 12, 14, 16, 18 – 21; Control sites: 8 – 10, 13, 15, 17, 22 and 23.

### 7.3 Results

The majority of trace metal concentrations in mussels collected from 15 of the 16 channel buoys (one buoy lacked mussels) were within the established national guidelines and consumable threshold levels for South Africa and other countries of the world (**Table 18**). However, as in the previous monitoring events this year, the median concentrations of mercury at impact and control sites continued to exceed the CSMP and the national maximum threshold (**Table 19**).

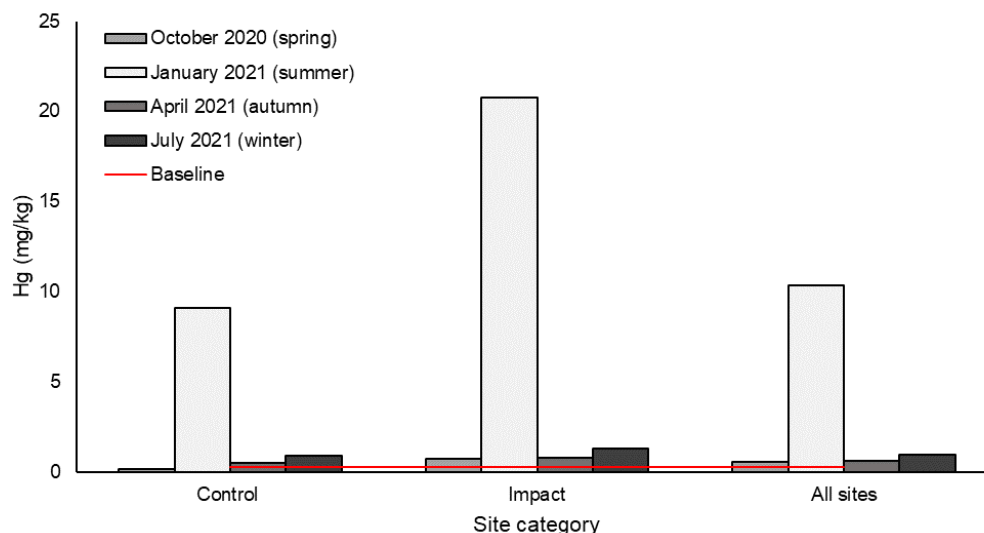
**Table 18:** Median concentrations of trace metals in *Perna perna* collected throughout the Port of Durban in winter 2021. Medians are given for all sites, impact (site 11, 12, 14, 16, 18-21) and control (site 8, 9, 10, 15, 17, 22). Values printed in red exceed the 90th percentile threshold of the CSMP. National Guideline levels are given for reference to other work. Note that the concentrations for mercury are given as part per million and not parts per billion. \*Cadmium levels below laboratory detection limits.

Metal (mg/kg)	90th percentile	National guideline level	Median (all sites)	Median (impact sites)	Median (control sites)
Aluminium	--	--	27.08	14.93	33.26
Arsenic	3.61	3	1.48	1.42	1.75
Cadmium	1.08	3	0*	0*	0*
Chromium	8.44	--	1.69	1.61	1.75
Cobalt	0.95	--	0.12	0.12	0.12
Copper	23.18	70	2.67	2.25	2.8
Iron	800.11	--	43.84	34.31	52.1
Lead	8.95	0.5	0.52	0.36	0.8
Manganese	134.75	--	14.07	11.94	18.17
Mercury	0.293	0.5	0.99	1.3	0.9
Nickel	5.6	--	0.72	0.69	0.75
Zinc	223.88	150	16.08	14.86	16.08

**Table 19:** Raw laboratory data of trace metal concentrations in brown mussels (*Perna perna*) collected during the winter 2021 survey from 15 channel buoys (control and impact sites) adjacent to the sandbanks in the Port of Durban. Concentrations that exceed the baseline guideline values are printed in red. Values marked with \* were below laboratory detection limits.

Trace Metal (mg/kg)	Control Sites							Impact Sites							
	Site 8	Site 9	Site 10	Site 13	Site 15	Site 17	Site 22	Site 11	Site 12	Site 14	Site 16	Site 18	Site 19	Site 20	Site 21
Aluminium	27.08	18.58	37.83	31.17	42.88	33.26	49.76	12.13	8.39	28.12	9.06	24.18	11.06	17.73	57.10
Arsenic	3.10	1.77	1.83	1.75	0.53	0.77	1.17	1.19	1.86	1.48	0.94	1.27	1.50	1.37	1.77
Cadmium	0.11	0.11	0.00	<17*	<17*	<17*	<17*	0.00	<17*	0.00	<17*	<17*	0.01	<17*	<17*
Copper	8.95	2.99	2.51	3.08	2.57	2.67	2.80	2.31	2.03	2.77	2.18	3.09	1.79	2.67	1.76
Cobalt	0.24	0.09	0.10	0.13	0.08	0.37	0.12	0.12	0.12	0.25	0.08	0.12	0.37	0.11	0.09
Chromium	2.31	2.18	1.75	1.58	1.51	1.65	1.97	1.52	1.41	2.40	1.41	1.47	1.78	1.69	1.80
Manganese	19.77	11.76	23.89	18.17	11.85	27.58	11.97	9.27	15.96	30.49	7.70	28.03	7.34	14.07	9.82
Nickel	1.55	0.59	0.73	0.75	0.58	0.98	0.84	0.60	0.68	1.03	0.62	0.72	0.60	1.68	0.70
Lead	2.46	2.23	0.24	0.41	0.55	1.00	0.80	0.13	0.20	<8*	0.68	0.52	1.04	0.13	<8*
Zinc	20.89	11.40	10.02	40.76	13.52	42.82	16.08	13.41	16.32	34.16	5.75	29.99	11.29	22.26	9.99
Mercury	0.90	0.85	0.94	1.57	0.75	1.51	0.86	1.60	1.24	1.35	1.70	0.99	4.19	0.45	0.70
Iron	63.58	38.97	54.63	43.84	35.29	52.05	69.24	24.44	28.64	46.58	25.99	46.44	36.23	32.38	49.03

In summer 2021, mercury concentrations were alarmingly high (**Figure 13**) and at levels that represented serious health risks for human and marine life. In subsequent sampling events (autumn and winter 2021), these concentrations were lower but still not below allowable national or local baseline thresholds. These concentrations have increased from autumn to winter. This therefore remains a matter of concern and requires increased management attention.



**Figure 13:** Median mercury concentrations (mg/kg) quantified from October 2020 (spring) to July 2021 (winter). Red line = baseline threshold value (0.29306 mg/kg).

## 7.4 Conclusion

Biomonitoring of mussels is an important tool for the assessment of pollution levels and indices of when action is required to maintain good water quality. All metals, apart from mercury, were below maximum baseline concentrations. These levels were slightly greater than those recorded in April and remain above the allowable threshold values and therefore still represents a cause for concern, especially as slight increases were recorded in winter. These concentrations remain at levels that pose health risks to animals and humans.

Bioaccumulation of trace metals in mussels take place over long periods, so the specific driver behind these elevated mercury concentrations will be difficult to pinpoint. Mercury can enter the marine environment through various sources, including agricultural run-off, waste from manufacturing electrical equipment, mine tailings and the burning of fossil fuels (DWAF 1995). The crude oil spill that occurred in the Umbilo river in October 2020 and/or the increased effluent and storm water runoff into the Port associated with the rainy season may be potential drivers. Nonetheless, this remains an important problem that needs to be urgently addressed by Port management authorities.



## 8 Fish

### 8.1 Introduction

Fish communities in the Port of Durban have been described as very diverse in past research conducted in the 1950s, where around 186 fish species were recorded (Day & Morgan, 1956). During this time, the most common species included *Terapon jarbua*, *Mugil cephalus*, *Liza dumerili*, *Ambassis dussumieri* and *Leiognathus equulus*. Many of these species are recognised as being dependent on estuaries, and the Bay was revealed as an important nursery areas for these and other economically important marine fish species. To this end, the shallow sandbanks and the little lagoon area specifically have been described as especially valuable nursery habitats (Cyrus & Forbes, 1996; Forbes & Demetriades, 2003).

More recent fish surveys in the Port by Angel and Clark (2008) and Newman et al. (2008) recorded far fewer species, at 29 and 34 species respectively. Most of the species either are listed as “Least Concern” on the IUCN Red List (2013) or have not been assessed. In these studies, the most abundant species have always been the Bald Glassy (*Ambassis dussumieri*), a small species that often dominates abundance but not biomass.

#### 8.1.1 Threshold limits set in the Central Sand Bank Mitigation Plan, 2017

The CSMP study stipulated various thresholds applicable to fish monitoring. According to this study, the median values of fish abundance, biomass and species richness are not to drop below the 80<sup>th</sup> percentile of the season-specific baseline median value (**Table 20**). If levels drop below limits specified, recovery of these faunal components in the Port may be delayed.

**Table 20:** Threshold (warning) levels to be adopted for fish abundance (no. ind./haul), biomass (g/haul) and species richness (no. species/haul) for the impact monitoring stations (adapted from Clark et al., 2017).

	Spring	Summer	Autumn	Winter
<b>Abundance</b>				
Median	15.2	31.6	408.7	255.3
80%	19.0	39.5	510.9	319.2
<b>Biomass</b>				
Median	175.2	252	711.15	2126.1
80%	219	315	888.9	2657.6
<b>Species richness</b>				
Median	4	4.4	5.2	4.8
80%	5.0	5.5	6.5	6.0

---

## 8.2 Methodology

### 8.2.1 Sampling protocol

The nearshore fish community in the Port of Durban was sampled using a beach seine net of 30 m length, 2 m depth and a stretched mesh size of 12 mm. Seine netting was conducted at 14 stations along the margins of the main sandbank areas in the Port (**Figure 14**). These sites are characterised as either impact or control sites in relation to the planned developments (**Table 21**). The net was deployed from a small fishing boat 30 – 50 m from the shore during daylight hours. All fish caught in the net at each station were identified, counted, weighed, measured and, where possible, returned to the estuary alive. All species caught were classified using an adaption of Whitfield's (1994) estuarine fish classification system. Four broad categories were used: marine, estuarine dependent, estuarine resident and freshwater species.

**Table 21:** Classification of impact and control sites for fish surveys in the Port of Durban.

Habitat	Site	Treatment
Central bank	6	Impact
Central bank	7	Impact
Central bank	8	Impact
Central bank	9	Impact
Central bank	10	Impact
Little lagoon	11	Impact
Little lagoon	12	Impact
Little lagoon	13	Control
Little lagoon	14	Control
Northern bank	19	Control
Northern bank	20	Control
Northern bank	21	Control
Northern bank	22	Control
Northern bank	23	Control



**Figure 14:** Location of fish sampling stations in the Port of Durban. Yellow pins = control sites, green pins = impact sites, black pins = additional sampling sites that will be included after the construction of the berths to monitor sandbank recovery.

## 8.2.2 Statistical Analyses

Multivariate analyses were conducted using PRIMER (v.6) where fish abundance data were subjected to a fourth root transformation to reduce the weight of abundant species and to achieve a balance of contribution between the rare and most common species. All analyses were performed using Bray-Curtis similarities of the fourth root abundance data. A PERMANOVA (Permutational Analysis of Variance) was used to determine if there are significant differences in the composition of fish communities found around impact and control sites. This test produces a p-value which, when smaller than 0.05 indicates a significant difference between the fish communities that inhabit the different sandbanks. Thereafter a SIMPER (Similarity Percentage) test was used to elucidate which species contributed the most to overall community similarity at impact and control sites.

## 8.3 Results

### 8.3.1 Key results

Very few fish were caught in winter 2021. The median biomass of 24 g for all sites was the lowest recorded in winter in the past few years of monitoring. Median abundance and species richness were similarly low and beneath the threshold minimum values stipulated for winter (**Table 22**).

**Table 22:** Median values of fish community dynamics at impact and control sites identified during the winter 2021 survey. Values given alongside 80th percentile thresholds stipulated in CSMP baseline study. Values printed in red violate CSMP threshold guidelines.

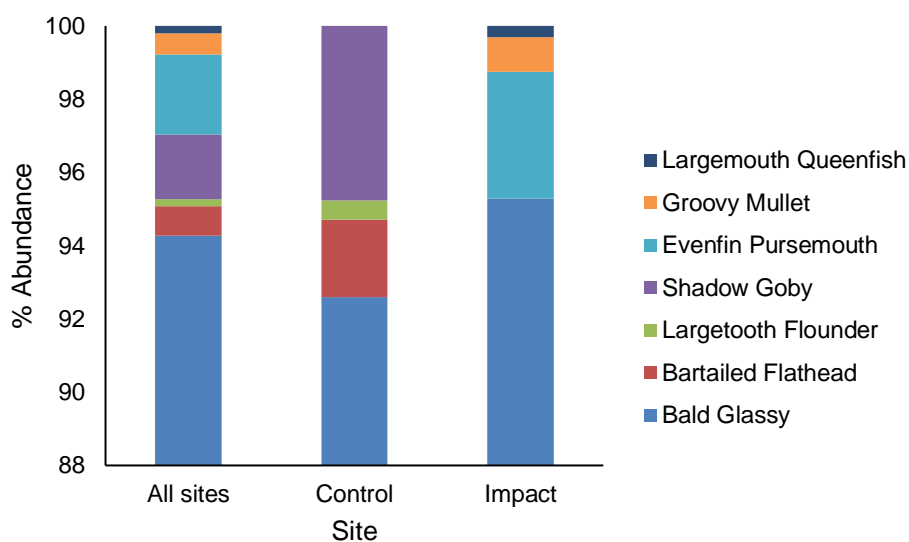
Parameter	80% limit (CSMP)	Median Overall	Median Impact	Median Control
Abundance	319.2	6	2	24
Biomass	2657.6	24	22.53	24
Species richness	6	1	1	1.5

### 8.3.2 Catch Composition

The catch composition comprised of 507 fish (~ 1 kg), representing 7 species. This is a substantial decrease from the 20 173 fish (~ 61.6 kg) and 13 species that were recorded in autumn. The total catch was, as in previous seasons, dominated by Bald Glassy (*Ambassis dussumieri*) that contributed 94.28% to the overall abundance and 61.61% to overall biomass. Other species detected include the Shadow Goby (*Yongeichthys nebulosus*) (1.7 % in number, 0.79% in mass), Bartailed Flathead (*Platycephalus indicus*) (0.79% in number, 3.9% in mass) and Groovy Mullet (*Liza dumerilii*) (0.59% in number, 24.8% in mass) (**Table 23, Table 24, Figure 15, Figure 16**).

**Table 23:** Abundance of fish species caught overall (all sites) and at control and impact sites in winter 2021.

Common name	Species	All sites	Control	Impact
Bald Glassy	<i>Ambassis gymnocephalus</i>	94.3	92.6	95.3
Bartailed Flathead	<i>Platycephalus indicus</i>	0.79	2.12	0
Largeetooth Flounder	<i>Pseudorhombus arsius</i>	0.2	0.53	0
Shadow Goby	<i>Yongeichthys nebulosus</i>	1.78	4.77	0
Evenfin Pursemouth	<i>Gerres methuen</i>	2.17	0	3.46
Groovy Mullet	<i>Liza dumerilii</i>	0.59	0	0.94
Largemouth Queenfish	<i>Scomberoides commersonnianus</i>	0.20	0	0.31

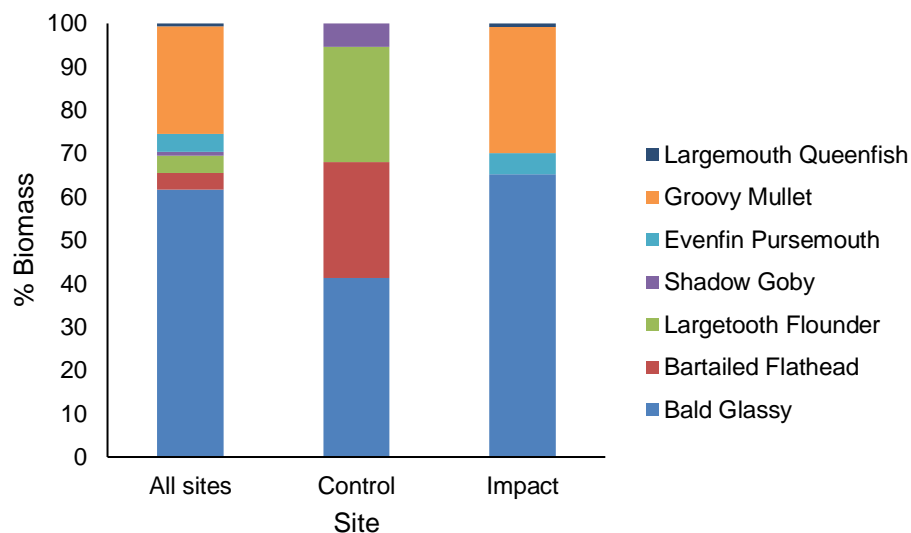


**Figure 15:** Relative abundance (%) of fish species caught overall (all sites) and at impact and control sites. Note that the y-axis starts at 88%.

Species that dominated by mass include Bald Glassy, Bartailed Flathead and Groovy Mullet (Table 24, Figure 16).

**Table 24:** Mass (g) of fish species caught overall (all sites) and at control and impact sites in winter 2021.

Common name	Species	All sites	Control	Impact
Bald Glassy	<i>Ambassis gymnocephalus</i>	61.61	41.3	65.16
Bartailed Flathead	<i>Platycephalus indicus</i>	3.97	26.67	0
Large-tooth Flounder	<i>Pseudorhombus arsius</i>	3.97	26.67	0
Shadow Goby	<i>Yongeichthys nebulosus</i>	0.8	5.33	0
Evenfin Pursemouth	<i>Gerres methuen</i>	4.2	0	4.89
Groovy Mullet	<i>Liza dumerilii</i>	24.8	0	29.13
Large-mouth Queenfish	<i>Scomberoides commersonnianus</i>	0.69	0	0.82



**Figure 16:** Relative mass (%) of fish species caught overall (all sites) and at impact and control sites.

### 8.3.3 Multivariate analyses and spatial patterns of diversity

Fish community composition differed significantly between impact and control sites (Pseudo- $F = 3.32$ ,  $df = 1$ ,  $p > 0.05$ ) and a subsequent SIMPER test revealed that the communities at impact sites were characterised by Evenfin Pursemouth while those at impact sites were characterised by Bald Glassy. Species richness and Shannon-Weiner  $H'$  indices for all sites surveyed are summarised below (**Table 25**).

**Table 25:** Species richness and Shannon-Weiner H' indices of fish communities surveyed at 14 monitoring stations within the Port of Durban in winter 2021.

Site number	Habitat	Richness	Diversity (H')
<b><i>Impact Sites</i></b>			
6	Central Bank	1	0
7	Central Bank	2	0.5
8	Central Bank	2	0.02
9	Central Bank	1	0
10	Central Bank	1	0
11	Little Lagoon	1	0
12	Little Lagoon	0	0
<b><i>Control Sites</i></b>			
13	Little Lagoon	2	0.69
14	Little Lagoon	2	0.26
19	Northern Bank	2	0.30
20	Northern Bank	1	0
21	Northern Bank	1	0
22	Northern Bank	3	0.54
23	Northern Bank	1	0

### 8.3.4 Estuarine Association in Fish Communities

Estuarine associations of marine and estuarine fish species have been well described by Whitfield (1994), especially species that live and breed in estuaries (referred to as estuarine resident species). Estuarine dependent species, on the other hand, are marine fishes that breed at sea but whose juveniles show varying degrees of dependency of estuaries. In winter 2021, catches were dominated by the estuarine resident species (mainly Bald Glassy).

## 8.4 Conclusion

Median fish abundance, biomass and species richness violated the CSMP guideline values. These values were some of the lowest recorded to date. There is a possibility that the water temperature on the date of sampling might have affected these results, as water temperature on the day of sampling was comparatively cooler (~14° C) than is typical of this habitat.

---

However, the values recorded are nonetheless concerning, especially as declining trends have been recorded over past sampling seasons.

As in previous seasons, Bald Glassy dominated catches, but even small numbers of this usually abundant species were recorded. Construction had not yet commenced at the time of this survey, so these data can be viewed as complimentary to the baseline data. Importantly, the fact that species richness continues to be low indicates that fish communities in the Port are at risk of losing important species and becoming homogenized.



---

## 9 Birds

### 9.1 Introduction

The natural habitats within Durban Bay have long been transformed and replaced by infrastructure associated with harbour developments. The original tidal flats, mangrove forests, and other littoral vegetation have vastly been reduced and replaced by open water areas and concrete berths. According to Allan et al. (1999), bird abundance and diversity dropped to 30% of that recorded in 1965 and avifaunal health of the Port was rated as poor in the 2011 National Biodiversity Assessment, due to substantial habitat loss and poor prey abundance and diversity (Driver et al., 2012).

Prior to construction, the aim of the avifaunal monitoring programme was to assess spatial and seasonal variability in avifaunal community structure. The monitoring must be continued during the construction phases to assess the impacts and effectiveness of the mitigation measures implemented during construction. This also forms part of the avifaunal monitoring programme recommended in the Bay of Natal Estuarine Management Plan (ERM/MER, 2012 and 2015), which proposes monthly surveys of birds on the intertidal, shallow subtidal, and mangrove habitats of the Bay.

### 9.2 Methodology

#### 9.2.1 Sampling protocol

The monitoring programme has been designed to align with the long-term monitoring already being conducted by the Durban Natural Science Museum (Allan et al., 1999; McInnes et al., 2005; Allan, 2012). The methodology followed that defined in the CSMP (Clark et al., 2017).

Five natural and semi-natural habitats have been identified in 20 sampling areas in Durban Bay (**Figure 17**), which will be monitored during the construction phase:

- Centre Bank and Little Lagoon (Bayhead North, MW West, MW East, CT West, CT East, FW North, FW South and Centre Bank Roost)
- Northern Sand Banks (Yacht Basin, Fish Wharf East and Fish Wharf West)
- Muddy/mangrove habitat (Sporting Bodies, Pelican Island, Bayhead West, Bayhead Central)
- Island View Sandbank
- Open Water (Harbour East, West, North and Harbour Mouth)



**Figure 17:** Aerial view of Durban Bay (Port of Durban) with 20 bird survey areas.

In the series of monitoring sections, individual birds of each species are counted and recorded by circumnavigating the inner periphery of the Port on a motorised vessel once per month at spring-low tide between 08h30 and 12h30. Bird counts are conducted by at least two observers with the aid of binoculars. To avoid double counting, observers count birds to the right of the vessel and flying overhead from the front only.

## 9.2.2 Statistical analyses

Data collected during June, July and August 2021 (winter) are included in this report. Data have been collated and used to characterise overall taxonomic composition, community structure, abundance, species richness and diversity, and compared to the baseline / preconstruction situation. Diversity indices were used as a measure of species richness and evenness to define community structure and diversity. The Shannon-Wiener diversity index ( $H'$ ) and the Pielou's evenness index ( $J'$ ) were calculated for each habitat type.

## 9.3 Results

### 9.3.1 Key results

Bird abundance, but not species richness, was above the minimum thresholds set for this period (**Table 26**). This was also the observed in previous seasons. The fact that species richness continues to be lower than the minimum thresholds set in the CSMP remains a matter

of concern. As the construction phase of this project has not yet commenced these declines are not related to project-specific impacts but rather most likely due to other anthropogenic drivers or natural fluctuations. The degree of plastic pollution in the Port remains a persistent threat to the health of bird communities as much of this litter accumulates on the sand banks that birds depend on. It affects feeding opportunities for birds both in the water (for piscivorous species) and on the sandbanks (for invertebrate feeders). Field workers are also experiencing increased difficulty with bird counts among the accumulating plastic litter on all of the sand banks that form part of the study area (**Figure 18**).

**Table 26:** Comparison with threshold levels of bird numbers and species richness recorded for Central Bank and Little Lagoon in the Durban harbour over the sampling period for all bird species. Values in red violate threshold limits.

	June	July	August
<b><i>CSMP threshold</i></b>			
<b>Total # species</b>	19	20	20
<b>Total # individuals</b>	324	336	317
<b><i>Jun '21 – Aug '21</i></b>			
<b>Total # species</b>	20	12	12
<b>Total # individuals</b>	443	357	927

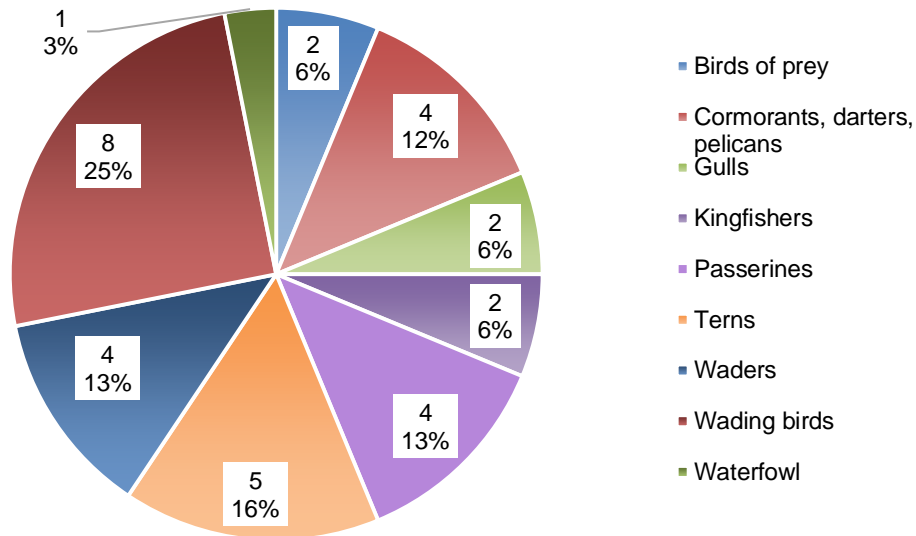


**Figure 18:** Accumulating plastic pollution on the Central Bank where monthly bird counts are conducted (January 2021).

### 9.3.2 Avian community composition

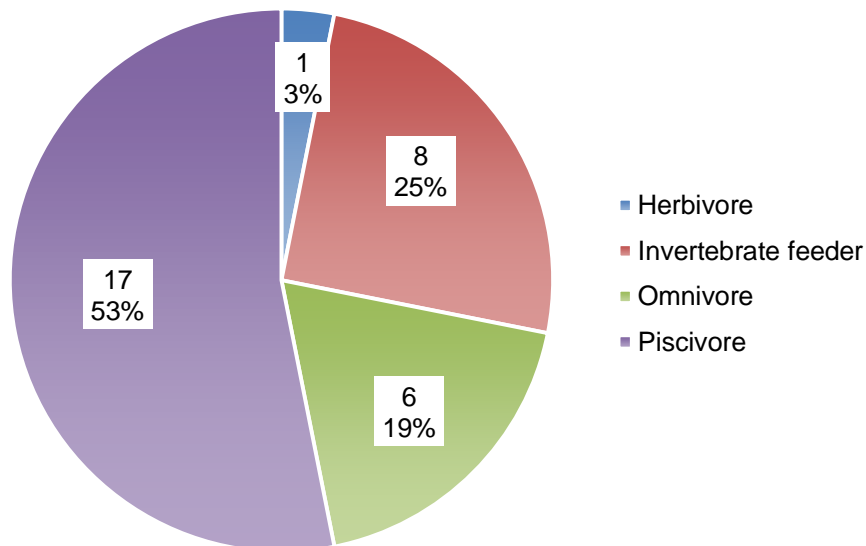
A total of 2 502 individual birds, representing 32 species were recorded during the three-month spring sampling period (June - August 2021). The largest proportion was counted at the Centre Bank and Little Lagoon (1 727 birds), which is in line with the overall findings of the baseline study. As in the baseline surveys, waders and wading birds comprised the largest proportion, consisting of four (13%) and eight species (25%), respectively (**Figure 19**). This was followed by terns comprising five species (16%).





**Figure 19:** Taxonomic composition (number and percentage of species) of bird communities recorded during monthly counts from June - August 2021.

In terms of feeding guilds, piscivores (fish eaters) and invertebrate feeders made up the largest proportion of bird species recorded over the three-month sampling period, each contributing 17 (53 %) and eight (36 %) species, respectively (**Figure 20**). This is also comparable to the baseline scenario where piscivores and invertebrate feeders comprised the largest proportion of birds observed over the sampling period.



**Figure 20:** Composition of feeding guilds (number and percentage of species) recorded during monthly counts from June - August 2021.

The avifauna typically found in Durban harbour can be divided into eight different taxonomic orders (**Table 27**). The group with the greatest species richness in this three-month period

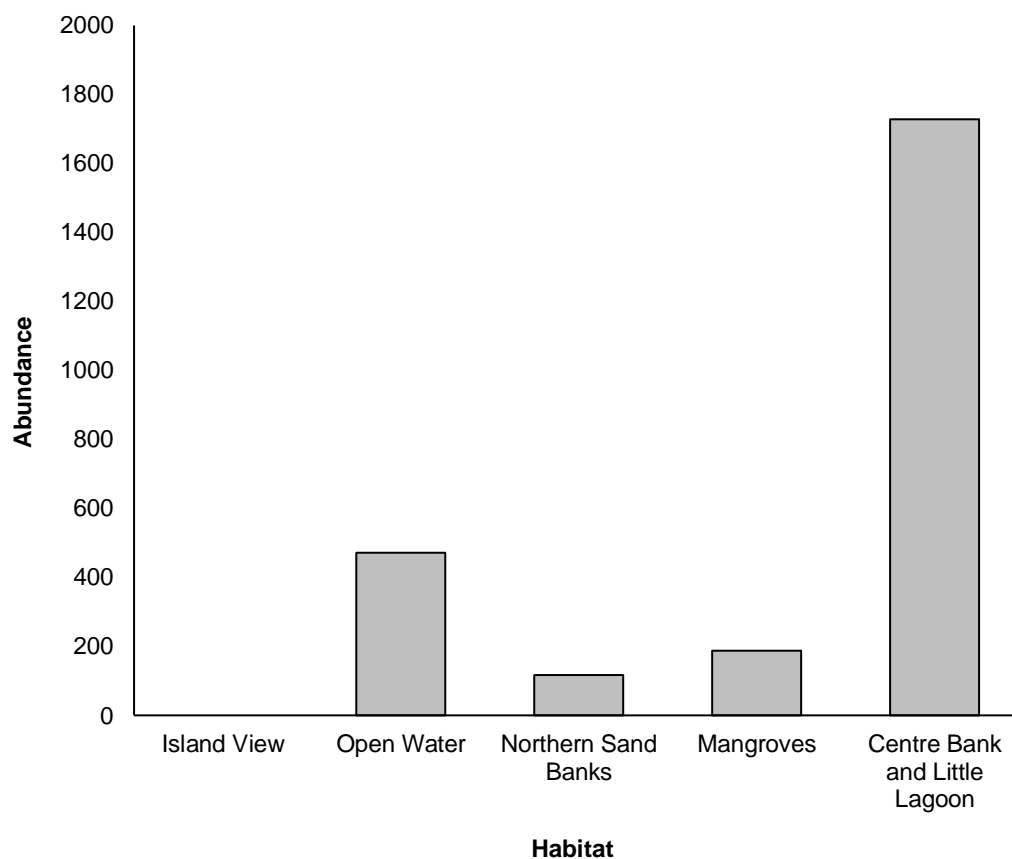
were the Charadriiformes (waders, gulls and terns) comprising 11 of the 32 species recorded (34.4 %). Within this group are the migratory waders and terns, which make up the highest number of migratory species (three species each) recorded in this sampling period. Migratory species made up 21.9 % of the total species recorded, which is typical during a winter period when some migratory species are absent.

**Table 27:** Taxonomic composition of common water-associated birds in the Port of Durban over the three-month sampling period (June - August 2021)

<b>Bird Group</b>	<b>Order</b>	<b>No. of SA Resident Species</b>	<b>No. of Migrant Species</b>
Birds of prey	Falconiformes	1	1
Cormorants, darters, pelicans	Pelecaniformes	4	
Gannets	Suliformes	0	
Gulls	Charadriiformes	2	
Kingfishers	Coraciiformes	2	
Passerines (swallows, martins, wagtails)	Passeriformes	4	
Terns	Charadriiformes	2	3
Waders	Charadriiformes	1	3
Wading birds (herons, egrets, ibises)	Ciconiiformes	8	
Waterfowl (ducks, geese)	Anseriformes	1	
<b>Total</b>		<b>25</b>	<b>7</b>

### 9.3.3 Avian abundance, species richness and diversity

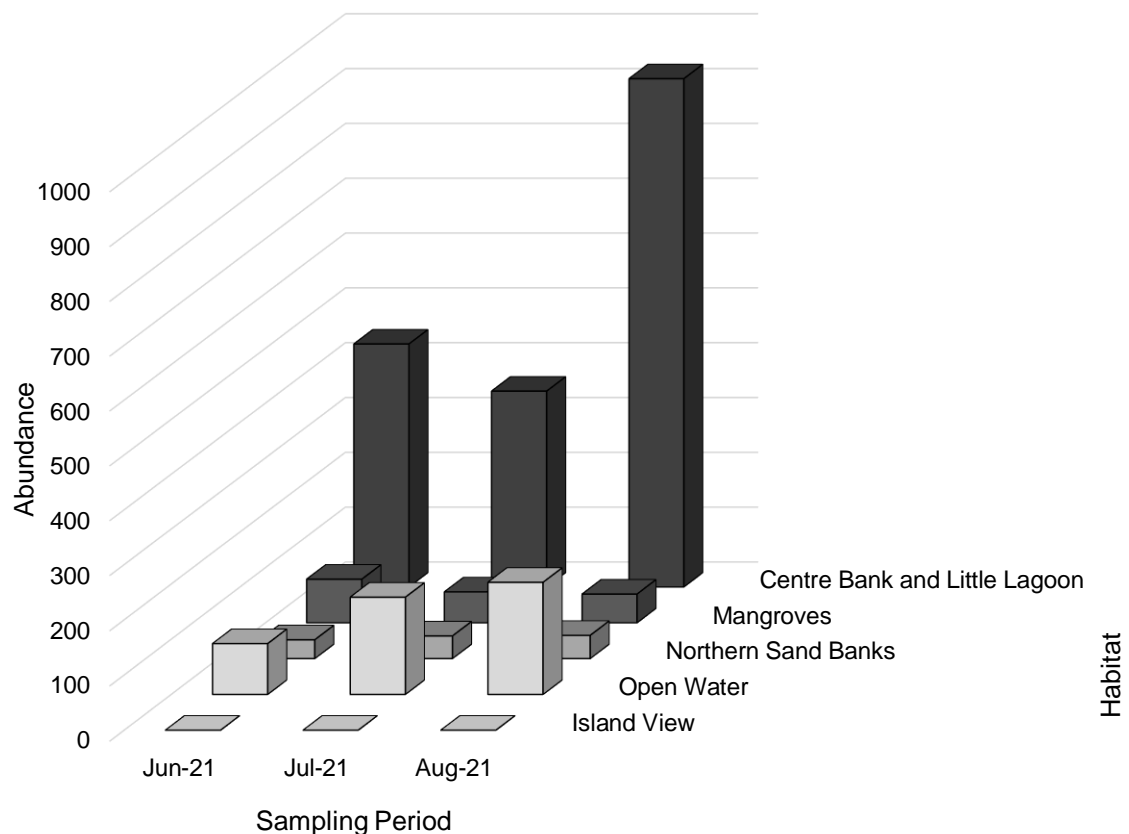
Of the five habitats identified within the Durban harbour, the Central Bank and Little Lagoon had the greatest number of birds (1 727) compared to Open Water (471), the Northern Bank (117), and Mangroves (187) in this three-month sampling period (**Figure 21**). The Central Bank and Little Lagoon had the greatest number of birds on a monthly basis, followed by the Mangroves then Open Water (**Figure 22**). No birds were recorded at Island View in this three-month period. Similar spatial patterns were observed during previous seasons and the baseline study, where the greatest numbers of birds were recorded from the Central Bank intertidal flats, and Island view had significantly lower bird abundance than all other sites. This is not surprising given that the Central Bank provides important feeding and roosting habitat for seabirds when exposed. It is also relatively isolated from harbour activities in comparison to other intertidal flats in the Port, so it may present the most undisturbed and attractive site for feeding and roosting birds. However, as emphasized in previous report, there is a continued concern that the degree of plastic pollution accumulating in this habitat may negatively affect avian communities.



**Figure 21:** Total bird abundance per habitat recorded during monthly counts from June – August 2021.

The number of birds recorded during the winter 2021 sampling period (2 502) was lower than the number recorded during spring and autumn sampling periods (3 539 and 2 967 birds,

respectively). This is likely reflective of departing migrant species from the area for winter (**Figure 22**). On a monthly basis, the Central Bank and Little Lagoon had the greatest number of birds, followed by Open Water and Mangroves (**Figure 22**).



**Figure 22:** Monthly bird abundance per habitat recorded from June – August 2021.

While species richness ( $S$ ) is purely a count of the number species present, diversity provides more information about the overall community composition. Species diversity incorporates relative species abundance and therefore provides information about rarity and commonness of species in a community (Stirling and Wilsey, 2001). Species diversity for bird communities in the Port of Durban is represented here by the Shannon Wiener Index ( $H'$ ), which calculates diversity based on both abundance and evenness of the species present. Evenness in this case is represented by Pielou's evenness index ( $J'$ ) and is constrained between zero and one. A higher number (i.e. closer to one) signifies a more even community, while a smaller number (i.e. closer to zero) suggests that the community is dominated by a few species (Stirling and Wilsey, 2001).

The greatest species richness was found at Open Water habitats ( $S = 25$ ) which also demonstrated the comparatively high diversity value ( $H' = 2.208$ ). Similar species richness and diversity was found at the Mangroves ( $S = 22$ ,  $H' = 2.133$ ) (**Table 28**). The comparatively high



diversity at the Mangroves may be explained by the fact that this is a protected area that offers a wide variety of habitats. In addition, the high diversity observed in the Open Water was likely driven by the fact that this habitat is used by piscivorous species for feeding and is a transition zone for birds moving between the other habitats. Certain species such as Egyptian Goose, Grey Heron, Little Egret and Blacksmith Lapwing also use the docks for resting, while White-breasted Cormorant are often found resting on the buoys in the harbour channels.

The abundances of species in the five main habitats were most evenly spread at the Mangroves and Northern Sand Banks (**Table 28**).

**Table 28:** Avifaunal species richness, diversity and evenness per habitat in the Durban harbour over the winter sampling period. No birds were recorded at Island View.

Habitat	Species richness (S)	Diversity (H')	Evenness (J')
Centre Bank and Little Lagoon	21	1.228	0.40
Mangroves	22	2.455	0.79
Open Water	25	2.133	0.66
Northern Sand Banks	15	2.139	0.79
Island View	0	n/a	n/a

## 9.4 Conclusion

The abundance of birds recorded during the winter 2021 sampling period was relatively lower than the number recorded during the previous two seasons. This is likely indicative of the departure of migrant species from the area for the winter. The Central Bank and Little Lagoon had the greatest bird abundance, while diversity was greatest at the Mangroves. The Central Bank provides important feeding and roosting (including shelter from the wind) habitat for seabirds when exposed. The Mangroves provide important natural habitat and is the only protected natural area in the Port. Possibilities for potential refuge sites, include the sandbanks at the Bayhead 'Natural Heritage' site, Fish Wharf opposite the Central Sandbank, and the smaller Yacht Basin, however to get full protection of these areas and to limit human disturbance will be difficult.

The sandbanks are not used for breeding by birds due to the transient nature of the intertidal flats (i.e. they become submerged at high tide every day), and many of the species utilising the sandbanks are non-breeding migrants. No nests or breeding birds are therefore at risk by the development at this time.

---

As found during the baseline surveys, waders and wading birds (mostly utilising the centre bank intertidal flats) were most abundant, more than half of the total number of species recorded during the three-month period. Within this group are the migratory waders and terns, which make up the highest number of migratory species recorded in the sampling period. Migratory species made up 21.9% of the total species recorded, highlighting the importance of managing the timing of disturbance to the centre bank during construction, with migratory season beginning in October and ending (based on previous studies) in April.

Piscivores (fish eaters) and invertebrate feeders made up the largest proportion of birds recorded in the three-month sampling period. The vast amount of litter in the harbour can impact on feeding opportunities for birds both in the water (for piscivorous species) and on the sandbanks (for invertebrate feeders). It is recommended that as an additional mitigation strategy for the approaching construction phase, the harbour is cleaned of litter on a regular basis, especially after heavy rainfall.

Overall abundance, but not species richness, was above the minimum thresholds stipulated in the CSMP. The consistently low species richness recorded in these seasonal surveys are a cause for concern and suggests that the Port may be at risk of losing some species for a prolonged period. As the construction phase for this project had not continued over the past year and a half, the lower values may be attributable to other anthropogenic or natural drivers. However, it is concerning that species richness was below allowable levels for another sampling season and indicates that intensified management / mitigation measures need to be implemented.

---

## 10 General Conclusion

Environmental aspects of this monitoring programme that failed to comply with limits set in the CSMP baseline study included the following:

- water turbidity (above maximum threshold),
- Total Organic Carbon content of sediments (below minimum threshold),
- macrofaunal abundance and species richness (below minimum thresholds)
- benthic microalgae (above maximum thresholds)
- mercury concentrations in mussels (above maximum threshold)
- abundance, biomass and species richness of fish communities (below minimum thresholds)
- bird species richness (below minimum thresholds).

The majority of these violations have persisted over several consecutive monitoring seasons and suggest that the overall ecological health of this system is poor. Ongoing chemical and plastic pollution are likely drivers of these observations and are concerning not only in terms of Port biodiversity but also for human health.

### 10.1 Water quality

Physico-chemical water quality parameters have been within ranges typical for marine environments. Typical seasonal decreases in water temperatures were observed. Salinity and turbidity levels were greater than those recorded in autumn, with turbidity values being nearly double that described in the baseline study. Minimal changes in pH and dissolved oxygen concentrations were observed.

### 10.2 Sediment characteristics

Medium to fine sand dominated intertidal and subtidal sediments and the total proportions of sand in the intertidal zone were slightly lower than recommended in the CSMP baseline study. Intertidal and subtidal TOC content were, as in previous seasons, below minimum allowable levels and is indicative of a nutrient-deficient system. The concentrations of all trace metals were within acceptable levels, including mercury that displayed concerning concentrations in summer.

### 10.3 Benthic macrofauna

A total of 38 intertidal and 27 subtidal macrofaunal species were identified in winter 2021. The median abundance and species richness in both zones are still lower than the minimum thresholds stipulated in the CSMP and have not been within the allowable levels for several seasons. While it is complicated to discern the exact drivers behind these concerning trends,

---

known factors such as the oil spill in October 2020 and severe and ongoing plastic pollution are likely candidates.

#### **10.4 Benthic microalgae**

Microalgae, measured in this case as the concentration of chlorophyll-a, is important in its role as primary producers and the concentration thereof play crucial roles in the flow of energy throughout the aquatic food web. As in previous seasons, intertidal habitats had greater concentrations of chl-a than subtidal habitats. The overall concentrations of chl-a concentrations increased from autumn and exceeded the maximum threshold concentrations for winter. In both tidal zones, this was driven by elevated concentrations at control sites on the northern bank.

#### **10.5 Biomonitoring using mussels (*Perna perna*)**

Assessment of trace metal concentrations in aquatic animals such as mussels are often used as an indication of the trace metal bioavailability in a habitat. In this regard, brown mussels (*Perna perna*) that grow on channel buoys across the Port of Durban are sampled. Mussels collected from 15 channel buoys had trace metal concentrations below the baseline thresholds for all metals apart from mercury. While mercury concentrations decreased from the exceptionally high levels recorded in summer, these concentrations remain above the allowable thresholds and can therefore continue to cause health problems for marine life and humans.

#### **10.6 Fish**

The abundance, biomass and species richness were below baseline threshold values and were the lowest recorded in the past few sampling seasons. While it is possible that cool water temperatures on the day of sampling played a role, the values recorded were severely low and represents cause for concern. This follows from previous seasons where the majority of community indices were below baseline threshold limits, and suggests that these communities are becoming increasingly homogenized.

#### **10.7 Birds**

Winter bird monitoring took place from June to August 2021. Avian species richness and was below the threshold minimum values in all three months, but abundance was above this threshold. In terms of abundance, this is a positive sign as abundance has not been above this threshold minimum for several seasons. However, the fact that species richness continues to decline remains concerning and suggests that there is a persistent decline in the overall diversity of avian communities across this area. There are various potential factors that may explain these declines, including the severe degree of plastic pollution in the Port of Durban. Marine birds often mistake plastic for food and feed it to their chicks that cannot digest plastic.

---

The Central Bank, a key roosting habitat for birds, has progressively become covered in plastic debris. This problem needs to be urgently addressed by Port management authorities.

---

## 11 References

- Agboola OA, Downs CT, O'Brien G. 2019. Macroinvertebrates as indicators of ecological conditions in the rivers of KwaZulu-Natal, South Africa. *Ecological Indicators* 106: 105465.
- Allan DG, Sinclair JC, Rossouw J. 1999. The waterbirds of Durban Bay: current status and historical trends. *Durban Museum Novitates* 24: 1-21.
- Allan DG. 2012. The waterbirds of Durban Bay: current status and historical trends. *Durban Museum Novitates* 35: 1-73.
- Allan DG, Sinclair JC, Rossouw J. 1999. The waterbirds of Durban Bay: current status and historical trends. *Durban Museum Novitates* 24: 1-21.
- Alves BGR, Guth AZ, Bicego MC, Gaeta SA, Sumida YG. 2014. Benthic community structure and organic matter variation in response to oceanographic events on the Brazilian SE inner shelf. *Continental Shelf Research* 85: 106-116.
- Anderson MJ. 2008. Animal-sediment relationships re-visited: Characterising species' distributions along an environmental gradient using canonical analysis and quantile regression splines. *Journal of Experimental Marine Biology and Ecology* 366(1): 16-27.
- Angel A, Clark BM. 2008. Port of Durban ballast water, flora and fauna survey. Anchor Environmental Consultants.
- Anthony EJ, & Héquette, A. 2007. The grain-size characterisation of coastal sand from the Somme estuary to Belgium: Sediment sorting processes and mixing in a tide-and storm-dominated setting. *Sedimentary Geology* 202(3): 369-382.
- Attrill MJ, Power M, Thomas RM. 1999. Modelling estuarine Crustacea population fluctuations in response to physico-chemical trends. *Marine Ecology Progress Series* 178: 89-99.
- Austen MC, Widdicombe S. 2006. Comparison of the response of meio- and macrobenthos to disturbance and organic enrichment. *Journal of Experimental Marine Biology and Ecology* 330(1): 96-104.
- Beck MW, Heck KL, Able KW, Childers DL, Eggleston DB. 2001. The identification, conservation, and management of estuarine and marine nurseries for fish and invertebrates. *Bioscience* 51: 633-641.
- Blackler JM, Forbes AT, Demetriades NT. 2004. The fish community of an intertidal sandbank and associated sub-tidal zone in Durban Harbour, South Africa: Implications of its removal and rehabilitation elsewhere in the harbour. In: Phang et al. (eds.) *Marine*

- 
- Science into the New Millennium: New Perspectives and Challenges. University of Malaya Maritime Research Centre.
- Bongers T, Ffris H. 1999. Nematode community structure as a bioindicator in environmental monitoring. *Trends in Ecology & Evolution* 14(6): 224-228.
- Brzeski V, Newkirk G. 1997. Integrated coastal food production systems—a review of current literature. *Ocean & Coastal Management* 34(1): 55-71.
- Buchman MF. 1999. NOAA screening quick reference tables.
- Caffrey JM. 2003. Production, respiration and net ecosystem metabolism in US estuaries. *In Coastal Monitoring through Partnerships* (pp. 207-219). Springer, Dordrecht.
- Cahoon LB, Cooke JE. 1992. Benthic microalgae production in the Onslow Bay, North Carolina, USA. *Marine Ecology Progress Series* 84(1): 185-196.
- Chadwick DB, Largier JL, & Cheng RT. 1996. The role of thermal stratification in tidal exchange at the mouth of San Diego Bay. *Coastal and Estuarine Studies*, 155-174.
- Christiansen KL, Raj Raman D, Anex RP. 2012. Predicting cost growth and performance of first-generation algal production systems. *Energy Policy* 51: 382–391.
- Clark BM, Harmer RW, Hutchings K, Brown EA, Massie VKR, Chidley C, Phamphe R. 2017 (October). Central Sandbank Mitigation Plan: Deepening, Lengthening and Widening of Berth 203 to 205, Pier 2, Container Terminal, Port of Durban. Prepared by Anchor Environmental and Nema Consulting.
- Clark BM, Harmer RW, Hutchings K, Brown EA, Massie VKR. 2016. Baseline Environmental Monitoring of Sand Banks in the Port of Durban. Technical Report No. 1486/4 prepared by Anchor Environmental Consultants for Nema Consulting and Transnet National Ports Authority.
- CSIR. 2016. Long-Term Ecological Monitoring for the Port of Durban Surveys made in 2015. CSIR Report.
- Cyrus DP, Forbes AT. 1996. Preliminary results on the role of KwaZulu-Natal harbours as nursery grounds for juveniles of selected marine organisms which utilise estuaries. *South African Journal of Wildlife Research* 26: 26-33.
- Dandonneau Y, Neveux J. 2002. An assessment of biases in fluorometric estimates of chlorophyll and pheophytin concentration in seawater, by reference to fluorescence spectral characteristics of main chlorophyllous pigments and large-scale sampling. *Instrumental Notes of IPSL* 18:1-19.
- Das S, Sahu BK. 2005. Interaction of pH with mercuric chloride toxicity to penaeid prawns from a tropical estuary, East Coast of India: enhanced toxicity at low pH. *Chemosphere* 58(9): 1241-1248.

- 
- David RA. 2006. A study of trace metals in estuarine sediment on the KwaZulu-Natal coast. Master of Science Dissertation, University of KwaZulu-Natal.
- Davis JP, Pitt KA, Connolly RM, Fry B. (2015). Food Webs.
- Day JH, Morgans JFC. 1956. The ecology of South African estuaries. Part 7. The biology of Durban Bay. *Annals of the Natal Museum* 13(3): 259-300.
- Deborde J, Anschutz P, Auby I, Glé C, Commarieu MV, Maurer D. ... Abril G. 2008. Role of tidal pumping on nutrient cycling in a temperate lagoon (Arcachon Bay, France). *Marine Chemistry* 109(1-2): 98-114.
- Department of Environmental Affairs (DEA), n.d. Review and Update of South Africa's National. Action List for the Screening of Dredged. Sediment Proposed for Marine Disposal.
- Department of Water Affairs and Forestry. 1995. South African Water Quality Guidelines for Coastal Marine Waters. Volume 1: Natural Environment.
- Desideri D, Meli MA, Roselli C, Feduzi L. 2009. A biomonitoring study: <sup>210</sup>Po and heavy metals in mussels. *Journal of Radioanalytical and Nuclear Chemistry* 279: 591-600.
- Doering PH. 1996. Temporal variability of water quality in the St. Lucia estuary, south Florida. *Journal of the American Water Resources Association* 32(6): 1293-1306.
- Driver A, Sink KJ, Nel JN, Holness S, Van Niekerk L, Daniels F, Jonas Z, Majiedt PA, Harris L, Maze K. 2012. National Biodiversity Assessment 2011: An assessment of South Africa's biodiversity and ecosystems, Synthesis Report, Pretoria: SANBI and DEA.
- Effendi H, Kawaroe M, Lestari DF. 2016. Ecological risk assessment of heavy metal pollution in surface sediment of Mahakam Delta, East Kalimantan *Procedia Environmental Sciences*: 33: 9.
- Ellingsen KE. 2002. Soft-sediment benthic biodiversity on the continental shelf in relation to environmental variability. *Marine Ecology Progress Series* 232: 15-27.
- ERM/MER 2012 and 2015. Bay of Natal Estuarine Management Plan Report prepared for eThekweni Municipality, Transnet National Ports Authority and the Department of Agriculture, Environmental Affairs and Rural Development.
- Fonseco EM, Fernandes JR, Lima LS, Delgado J, Correa TR, Costa PMS, Baptista Neto A, Aguiar VMC. 2020. Effects of dredged sediment dumping on trace metals concentration and macro benthic assemblage at the continental shelf adjacent to a tropical urbanized estuary. *Ocean and Coastal Management* 196: 105299.
- Forbes A, Demetriades N. 2003. Ecology of the "Little Lagoon Area" of Durban Bay. Report prepared for the National Port Authority by Marine & Estuarine Research.



- 
- Forbes AT, Demetriades NT, Cyrus DP. 1996. Biological significance of harbours as coastal habitats in KwaZulu-Natal, South Africa. *Aquatic Conservation: Marine and Freshwater Ecosystems* 6: 331-341.
- França S, Vasconcelos RP, Tanner S, Máguas C, Costa MJ, Cabral HN. 2011. Assessing food web dynamics and relative importance of organic matter sources for fish species in two Portuguese estuaries: a stable isotope approach. *Marine Environmental Research* 72(4): 204-215.
- Gaston GR, Young JC. 1992. Effects of contaminants of macrobenthic communities in the upper Calcasieu Estuary, Louisiana. *Bulletin of Environmental Contamination and Toxicology* 49(6): 922-928.
- Goher ME. 2014. Metal pollution assessment in the surface sediment of Lake Nasser, Egypt. *Egyptian Journal of Aquatic Research* 40: 12.
- Goodnight CJ. 1973. The use of aquatic macroinvertebrates as indicators of stream pollution. *Transactions of the American Microscopical Society* 1-13.
- Gray JS, Wu RS, Or YY. 2002. Effects of hypoxia and organic enrichment on the coastal marine environment. *Marine Ecology Progress Series* 238:249–279.
- Guastella LM. 1994. A quantitative assessment of recreational angling in Durban Harbour, South Africa. *South African Journal of Marine Science* 14(1): 187-203.
- Halpern BS, Walbridge S, Selkoe KA, Kappel CV, Micheli F, D'agrosa C, Fujita R. 2008. A global map of human impact on marine ecosystems. *Science* 319(5865): 948-952.
- Hauer FR, Resh VH. 2017. Macroinvertebrates. In *Methods in Stream Ecology*, Volume 1 (pp. 297-319). Academic Press.
- Hay DG, Forbes AT, Cyrus DP. 1995. The biological status of the ports of Natal and their role as alternative nursery grounds for juveniles of marine organisms normally occurring in estuaries. In: Hay D.G., Breen C.M., (eds). *Disturbance and evolution as factors influencing the structure and functioning of estuaries*. Investigational report for the Institute of Natural Resources of South Africa, Pietermaritzburg, IR107: 80-96.
- Hellweger FL, Schlosser P, Lall U, Weissel JK. 2004. Use of satellite imagery for water quality studies in New York Harbor. *Estuarine, Coastal and Shelf Science* 61(3): 437-448.
- Hindmarch JA, Griffiths AL, Officer AK, Wittneben G. 2008. Durban Harbour tunnel-first use of a slurry tunnel boring machine in South Africa. *Journal of the Southern African Institute of Mining and Metallurgy* 108(1): 1-8.
- Jones AB, O'Donohue MJ, Udy J, Dennison WC. 2001. Assessing ecological impacts of shrimp and sewage effluent: biological indicators with standard water quality analyses. *Estuarine, Coastal and Shelf Science* 52(1): 91-109.

- 
- Kamer K, Fong P. 2000. A fluctuating salinity regime mitigates the negative effects of reduced salinity on the estuarine macroalga, *Enteromorpha intestinalis* (L.) link. *Journal of Experimental Marine Biology and Ecology* 254(1): 53-69.
- Karr JR. 1993. Protecting ecological integrity: an urgent societal goal. *Yale J. Int'l L.*, 18, 297.
- Klapow LA, Lewis RH. 1979. Analysis of toxicity data for California marine water quality standards. Water Pollution Control Federation: 2054-2070.
- Kljaković-Gašpić Z, Odžak N, Ujević I, Zvonarić T, Horvat M, Barić A. 2006. Biomonitoring of mercury in polluted coastal area using transplanted mussels. *Science of the total environment*, 368(1), 199-209.
- Kljaković-Gašpić Z, Herceg-Romanić S, Kožul D, Veža J. 2010. Biomonitoring of organochlorine compounds and trace metals along the Eastern Adriatic coast (Croatia) using *Mytilus galloprovincialis*. *Marine Pollution Bulletin* 60(10): 1879-1889.
- Lee SW, Park G, Choi KH. 2019. Biomass of plankton and macrobenthos and benthic species diversity in relation to environmental gradients in a nationwide coastal survey. *Regional Studies in Marine Science* 26: 100502.
- Levit SM. 2010. A literature review of effects of cadmium on fish.
- Lopez G, Taghon G, Levinton J (eds). 1989. Ecology of marine deposit feeders. *Lecture Notes on Coastal and Estuarine Studies* 31. Springer, New York.
- MacIntyre HL, Geider RJ, McKay RM. 1996. Photosynthesis and regulation of rubisco activity in net phytoplankton from delaware bay<sup>1</sup>. *Journal of Phycology* 32(5): 718-731.
- Madeira D, Narciso L, Cabral HN, Vinagre C, Diniz MS. 2013. Influence of temperature in thermal and oxidative stress responses in estuarine fish. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology* 166(2): 237-243.
- McInnes AM, Allan DG, Malan G. 2005. Key sites for the conservation of waterbirds (Aves) in Durban Bay, South Africa. *Durban Mus. Novit*, 30, pp.1-14.
- MER/ERM. 2012. Development of the Bay of Natal Estuarine Management Plan: Situation Assessment. Report prepared for eThekweni Municipality, Transnet National Port Authority and the Department of Agriculture, Environmental Affairs and Rural Development.
- Moore KA, Wetzel RL, Orth RJ. 1997. Seasonal pulses of turbidity and their relations to eelgrass (*Zostera marina* L.) survival in an estuary. *Journal of Experimental Marine Biology and Ecology* 215(1): 115-134.
- N-Uptake A. (1999). Primary production by phytoplankton and microphytobenthos in estuaries. *Estuaries* 29: 93.

- 
- Newman B, Weerts S, Porter S, Mackay, F. 2008. Specialist studies for Transnet capital expansion program, Durban Bay. Ecosystems Competency, Council for Scientific and Industrial Research.
- Newman B, Weerts S, Porter S, Mackay F. 2008. Specialist studies for Transnet capital expansion program, Durban Bay. Ecosystems Competency, Council for Scientific and Industrial Research.
- O'Brien A, Townsend K, Hale R, Sharley D, Pettigrove V. 2016. How is ecosystem health defined and measured? A critical review of freshwater and estuarine studies. *Ecological Indicators* 69: 722-729.
- O'Connor TP. 2004. The sediment quality guideline, ERL, is not a chemical concentration at the threshold of sediment toxicity. *Marine Pollution Bulletin* 49(5-6): 383-385.
- Pacheco AS, Laudien J, Thiel M, Oliva T, Arntz W. 2010. Succession and seasonal variation in the development of sub-tidal macrobenthic soft-bottom communities off northern Chile. *Journal of Sea Research* 64: 180-189.
- Paerl HW, Pinckney JL. 1996. A mini-review of microbial consortia: their roles in aquatic production and biogeochemical cycling. *Microbial Ecology* 31(3): 225-247.
- Pearson TH, Rosenberg R. 1978. Macrobenthic succession in relation to organic enrichment and pollution of the marine environment. *Oceanography and Marine Biology Annual Review* 16: 229 - 311.
- Phillips DJH. 1980. *Quantitative Aquatic Biological Indicators: Their Use to Monitor Trace Metal and Organochlorine Pollution*. London: Applied Science Publishers.
- Phillips DJH. 1995. The chemistries and environmental fates of trace metals and organochlorines in aquatic ecosystems. *Marine Pollution Bulletin* 31(4-12): 193-200.
- Phillips DJH, Rainbow PS. 1993. *Biomonitoring of Trace Aquatic Contaminants*. Applied Science Publishers, Barking.
- Pillay S, Mapukata T, Connell A, Blair A, David R, Mudaly R, Manqele Z, Mthethwa P, Delport B, Dloma K, Ramathatsoenyane L, Ncgobo B, Mdluli O, Singh A. 2003. Water Quality Monitoring Program, Port of Durban, 2002-2003, CSIR. Final Report.
- Pillay D. 2002. *The macrobenthos of the Little Lagoon, Durban Bay (Doctoral dissertation)*.
- Pillay D, Perissinotto R. 2008. The benthic macrofauna of the St. Lucia Estuary during the 2005 drought year. *Estuarine, Coastal and Shelf Science* 77(1): 35-46.
- Pinckney J, Zingmark RG. 1993. Biomass and production of benthic microalgal communities in estuarine habitats. *Estuaries* 16(4): 887-897.
- Pinckney J, Zingmark RG. 1993. Biomass and production of benthic microalgal communities in estuarine habitats. *Estuaries* 16(4): 887-897.

- 
- Pinto R, Patricio J, Baeta A, Fath BD, Neto JM, Marques JC. 2009. Review and evaluation of estuarine biotic indices to assess benthic condition. *Ecological Indicators* 9(1): 1 - 25.
- Powell B, Martens M. 2005. A review of acid sulfate soil impacts, actions and policies that impact on water quality in Great Barrier Reef catchments, including a case study on remediation at East Trinity. *Marine Pollution Bulletin* 51(1): 149-164.
- R Core Team. 2020. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>.
- Rainbow PS. 1995. Biomonitoring of heavy metal availability in the marine environment. *Marine Pollution Bulletin* 31: 183-192.
- Rainbow PS. 2006. Biomonitoring of trace metals in estuarine and marine environments. *Australasian Journal of Ecotoxicology* 12: 107-122.
- Ray GL. 2000. Infaunal assemblages on constructed intertidal mudflats at Jonesport Maine (USA). *Marine Pollution Bulletin* 40: 118–1200.
- Reddi KR, Jayaraju N, Suryakumar I, Sreenivas K. 1993. Tidal fluctuations in relation to certain physic-chemical parameters in Swarnamukhi river estuary, east coast of India. NISCAIR-CSIR, India.
- Riera R, Tuya F, Ramos E, Rodriguez M, Monterroso O. 2012. Variability of macrofaunal assemblages on the surroundings of a brine disposal. *Desalination* 291: 94 - 100.
- Riera R, Tuya F, Rodriguez M, Monterroso O, Ramos E. 2013. Confounding response of macrofauna from a confluence of impacts: brine and sewage pollution. *Acta Oceanologica Sinica* 32(10): 74 - 81.
- Roberts DA, Johnston EL, Knott NA. 2010. Impacts of desalination plant discharges on the marine environment: A critical review of published studies. *Water Research* 44(18): 5117-5128.
- Ryu J, Khim JS, Kang SG, Kan D, Lee C, Koh C. 2011. The impact of heavy metal pollution gradients in sediments on benthic macrofauna at population and community levels. *Environmental Pollution* 159(10): 2622-2629.
- Saunders JE, Al Zahed KM, Paterson DM. 2007. The impact of organic pollution on the macrobenthic fauna of Dubai Creek (UAE). *Marine Pollution Bulletin* 54(11): 1715-1723.
- Seuront L, Leterme C. 2006. Microscale patchiness in microphytobenthos distributions: evidence for a critical state. In: Kromkamp JC, de Brouwer J, Blanchard GF, Forster RM & Créach V (eds.), *Functioning of microphytobenthos in estuaries*. The Academy of Arts and Sciences, Amsterdam/The University of Chicago Press, Chicago, 165-183.
- Snelgrove PV, Butman CA. 1994. Animal-sediment relationships revisited: cause versus effect. *Oceanography and Marine Biology Annual Review* 32: 111-117.

- 
- Snow GC, Taljaard S. 2007. Water quality in South African temporarily open/closed estuaries: a conceptual model. *African Journal of Aquatic Science* 32(2): 99-111.
- Sponseller RA, Heffernan JB, Fisher SG. 2013. On the multiple ecological roles of water in river networks. *Ecosphere* 4(2): 1-14.
- Stevenson RJ, Pan Y. 1999. Assessing environmental conditions in rivers and streams with diatoms. *The diatoms: applications for the environmental and earth sciences* 1(4).
- Stirling G, Wilsey B. 2001. Empirical Relationships between Species Richness, Evenness, and Proportional Diversity. *The American Naturalist* 158(3): 286–299.
- Streever WJ. 2000. *Spartina alterniflora* on dredged material: a critical view of the ongoing debate over success. *Wetlands Ecology and Management* 8: 295-316.
- Summers JK, Wade TL, Engle VD, Malaeb ZA. 1996. Normalization of metal concentrations in estuarine sediments from the Gulf of Mexico. *Estuaries and Coasts* 19(3): 581-594.
- Tengberg A, Almroth E, Hall P. 2003. Resuspension and its effects on organic carbon recycling and nutrient exchange in coastal sediments: in situ measurements using new experimental technology. *Journal of Experimental Marine Biology and Ecology* 285: 119-142.
- Turpie JK, Clark BM. 2007. The health status, conservation importance, and economic value of temperate South African estuaries and development of a regional conservation plan. Anchor Environmental Consultants (Pty) Ltd, Report to Cape Nature.
- Varadharajan D, Soundarapandian P, Pushparajan N. 2013. Effect of physico-chemical parameters on crab biodiversity. *Journal Marine Science and Research Development* 3: 116.
- Wang S, Jia Y, Wang S, Wang X, Wang H, Zhao Z, Liu B. 2010. Fractionation of heavy metals in shallow marine sediments from Jinzhou Bay, China. *Journal of Environmental Sciences* 22: 23-31.
- Warwick RM. 1993. Environmental impact studies on marine communities: pragmatical considerations. *Australian Journal of Ecology* 18:63-80.
- Watson EB, Pasternack GB, Gray AB, Goñi M, Woolfolk AM. 2013. Particle size characterization of historic sediment deposition from a closed estuarine lagoon, Central California. *Estuarine, Coastal and Shelf Science* 126: 23-33.
- Weerts SP. 2010. Elevation and bathymetric survey of the Port of Durban sandbanks post entrance widening, with a desktop assessment of potential ecological implications. CSIR Report CSIR/NRE/CO/IR/2010/0074/C, Durban.

- 
- Weinstein MP, Litvin SY, Guida VG. 2005. Considerations of habitat linkages, estuarine landscapes, and the trophic spectrum in wetland restoration design. *Journal of Coastal Research*: 51-63.
- Whitfield AK. 1998. *Biology and Ecology of Fishes in Southern African Estuaries*. Ichthyological Monographs of the J.L.B. Smith Institute of Ichthyology 2: 1-223.
- Whitney D, Darley W. 1983. Effect of light intensity upon salt marsh benthic microalgal photosynthesis. *Marine Biology* 75: 249-252.
- Wolfstein K, Stal LJ. 2002. Production of extracellular polymeric substances (EPS) by benthic diatoms: effect of irradiance and temperature. *Marine Ecology Progress Series* 236: 13-22.

# Document Control and Disclaimer

FORM IP180\_B



<b>CLIENT</b>	:	Transnet Group Capital	
<b>PROJECT NAME</b>	:	Transnet Berth 203 to 205 ECO	<b>PROJECT No.</b> : GE38097
<b>TITLE OF DOCUMENT</b>	:	Winter Monitoring Report 2021	
<b>ELECTRONIC LOCATION</b>	:	\\Durban-5\projects\J38097-Transnet Berths ECO\03_Project Management Plan Design\G_Document Management - Reports\Monitoring Data & Reports\2021\Winter Report	

Approved By		Reviewed By	Prepared By
Project Executive			
<b>ORIGINAL</b>	NAME W. Fyvie	NAME W. Fyvie, M. Muir, Dr L. Skein	NAME Dr Lisa Skein, Charl Kruger, Kolawole Akinoshun, Robyn Phillips
DATE <b>02/09/2021</b>	SIGNATURE	SIGNATURE	SIGNATURE

This report, and information or advice, which it contains, is provided by GIBB (or any of its related entities) solely for internal use and reliance by its Client in performance of GIBB's duties and liabilities under its contract with the Client. Any advice, opinions, or recommendations within this report should be read and relied upon only in the context of the report as a whole. The advice and opinions in this report are based upon the information made available to GIBB at the date of this report and on current South African standards, codes, technology and construction practices as at the date of this report. Following final delivery of this report to the Client, GIBB will have no further obligations or duty to advise the Client on any matters, including development affecting the information or advice provided in this report. This report has been prepared by GIBB in their professional capacity as Consulting Engineers. The contents of the report do not, in any way, purport to include any manner of legal advice or opinion. This report is prepared in accordance with the terms and conditions of the GIBB contract with the Client. Regard should be had to those terms and conditions when considering and/or placing any reliance on this report. Should the Client wish to release this report to a Third Party for that party's reliance, GIBB may, at its discretion, agree to such release provided that:

- GIBB's written agreement is obtained prior to such release, and
- By release of the report to the Third Party, that Third Party does not acquire any rights, contractual or otherwise, whatsoever against GIBB and GIBB, accordingly, assume no duties, liabilities or obligations to that Third Party, and
- GIBB accepts no responsibility for any loss or damage incurred by the Client or for any conflict of GIBB interests arising out of the Client's release of this report to the Third Party.